

Welcome!

Open Geneious (or your software of choice for viewing, annotating and editing sequences)

Open the workshop webpage: **<https://www.allencell.org/scge-workshop-2019.html>**

If you had any trouble completing the downloads of the NUP153 or mEGFP sequences, we are available to help you now!

Internet access:

“Institute Guest” pw: sounders



Stem cells and Gene editing workshop

3/27/2019

Ru Gunawardane
Director, Stem cells and Gene editing



ALLEN INSTITUTE *for*
CELL SCIENCE

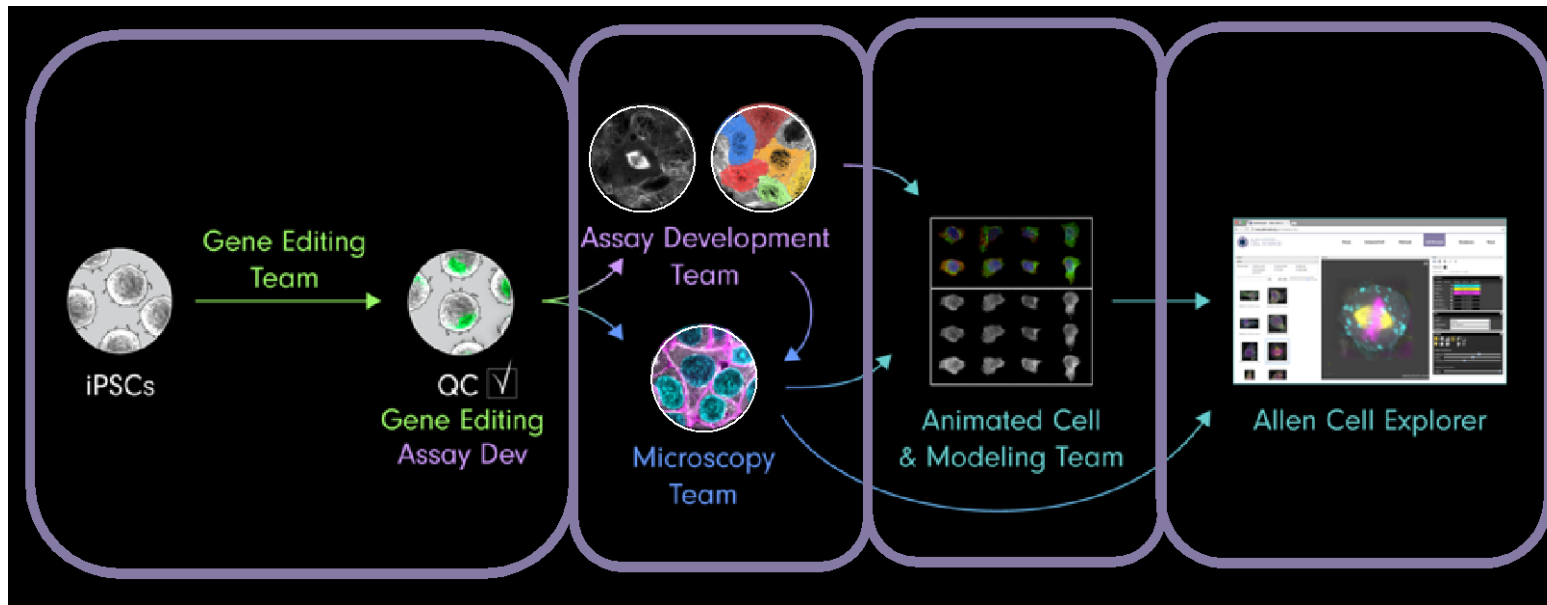
Allen Institute for Cell Science

Institute Mission

- Understand and predict cellular behaviors in normal and pathologic contexts
- Develop dynamic, visual data on cellular organization and activities
- Share our data and tools with the research community

Institute values

- Team science
- Open science
- Community empowerment



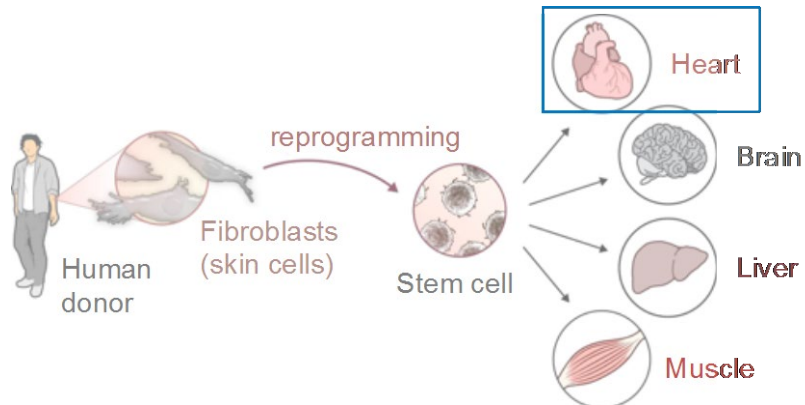
- Research program launched in January, 2016
- Highly multi-disciplinary team of ~65



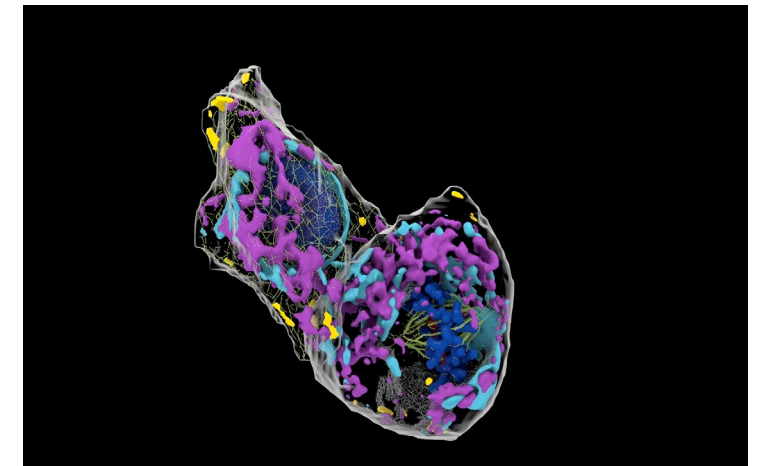
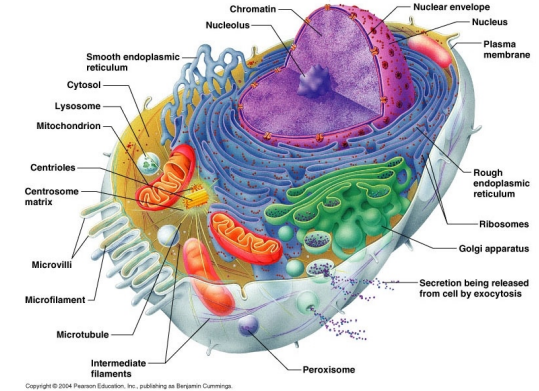
Creating a stem cell state map with gene edited hiPSCs and live imaging

- *Physical organization of the cell – “google” map of the cell*
- *Study how cells change across states (division, differentiation, disease)*

iPSCs as a cell model



- Human
- Diploid
- Non-transformed
- Proliferative
- Pluripotent



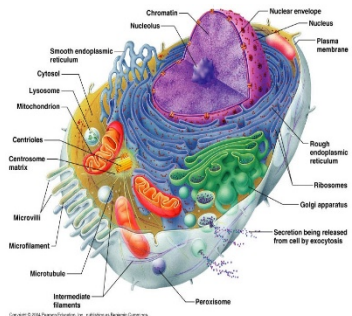
Data driven, integrated model of a cell

Critical points and rationale:

- iPSCs from a healthy donor - to understand baseline and variance
- Live imaging - capture dynamics and avoid artifacts of fixation and staining
- Endogenous tagging - Normal regulation of tagged structure
- High level of QC - data used for creating baseline models of cell behavior



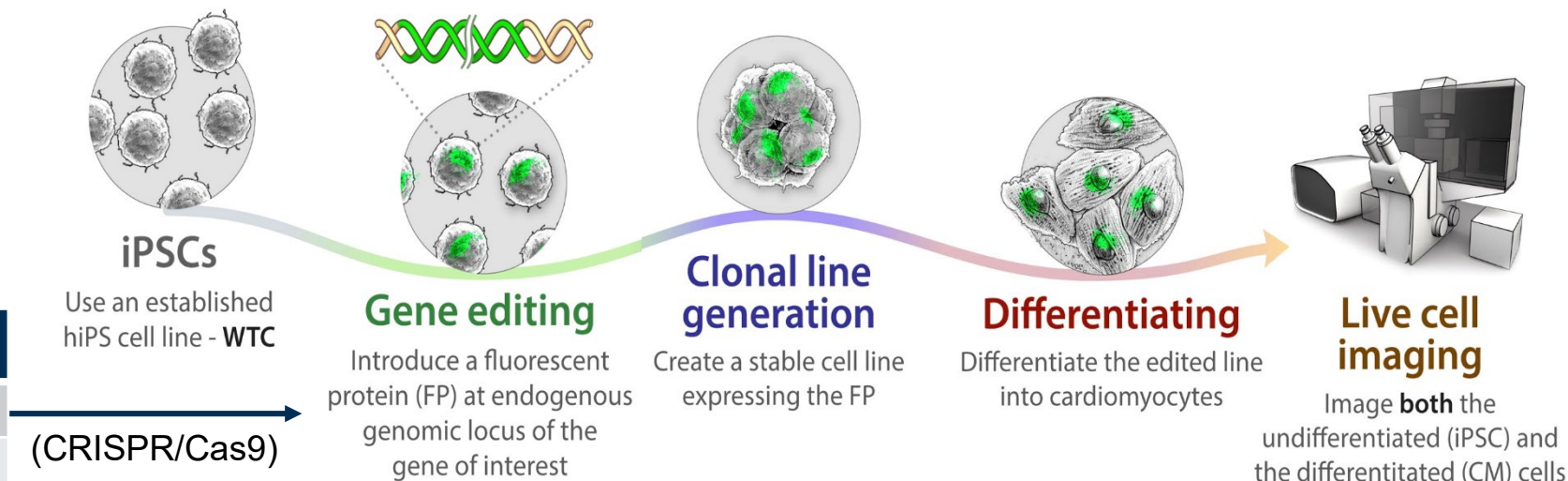
Creating a collection of FP tagged hiPSC lines representing major structures of the cells



Working with experts to select key proteins to tag and how to tag them



	Structure	Gene/Protein
1	Adhesions	Paxilin
2	Microtubules	Alpha tubulin
3	Nucleus	LaminB1
4	Mitochondria	Tom20
5	Cell-cell junctions	Desmoplakin
6	Actin	Beta actin
7	ER	Sec61B
8	Nucleolus	Fibrillarin
9	Golgi	ST6Gal1
10	Centrosome	Centrin2



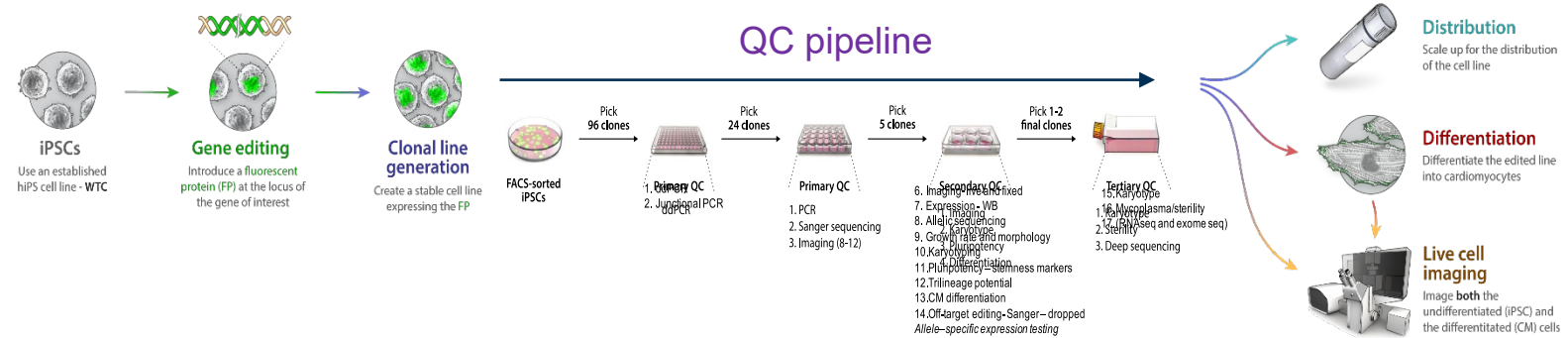
- Generate a collection of high quality clonal FP-tagged iPSC lines
- Develop editing strategy, workflows and QC methods
- Share cell lines, plasmids, images, and methods with the community



The Allen Cell and Plasmid Collections

	Structure	Gene/Protein	FP	Status
1	Adhesions	Paxilin	mEGFP	Available
2	Microtubules	Alpha tubulin	mEGFP	Available
3	Nuclear lamin	LaminB1	mEGFP	Available
4	Mitochondria	Tom20	mEGFP	Available
5	Cell-cell junctions	Desmoplakin	mEGFP	Available
6	Actin	Beta actin	mEGFP	Available
7	ER	Sec61B	mEGFP	Available
8	Nucleolus	Fibrillarin	mEGFP	Available
9	Myosin	Myosin IIB	mEGFP	Available
10	Tight junctions	ZO-1	mEGFP	Available
11,12	Cytoplasm	Safe harbor	mEGFP	Available
13	Centrosome	Centrin2	mTagRFPt	Available
14	Microtubules	Alpha tubulin	mTagRFPt	Available
15,16	Golgi	ST6Gal1	mEGFP	Available
17	Lysosome	LAMP1	mEGFP	Available
18	Autophagosomes	LC3	mEGFP	Available
19,20	Endosome	Rab5	mEGFP	Available
21	Peroxisomes	PMP34	mEGFP	Available
22	Plasma membrane	Safe harbor-CAAX	mTagRFPt	Available
23	Gap junctions	Connexin43	mEGFP	Available
24	Nucleolar	NPM1	mEGFP	Available
25	Nuclear pore	NUP153	mEGFP	Available
26	Histones	H2B	mEGFP	Available
27	Beta catenin	CTNNB1	mEGFP	Available
28	Phase transition	FUS	mEGFP	Available
29	Nuclear cohesin	SMC1A	mEGFP	Available
30	ER/Nuclear lamin	Sec61B/LMN1	mEGFP, mTagRFPt	Available
31	CM- Thin filament	ssTnni1	mEGFP	Available
32	CM- Thick filament -	MYL7 (early)	mEGFP	Available
33	CM- M-disc	Titin	mEGFP	Available
34	CM- SR	ATP2A2	mEGFP	Available
35	CM- Thick filament -	MYL2 (late)	mEGFP	Available
36	CM- Z-disc	ACTN2	mEGFP	Available
37	Transcription factor	Sox2	mEGFP	Pending
38	Nucleolar dual	FBL/NPM1	mEGFP	Pending

4-6 month process



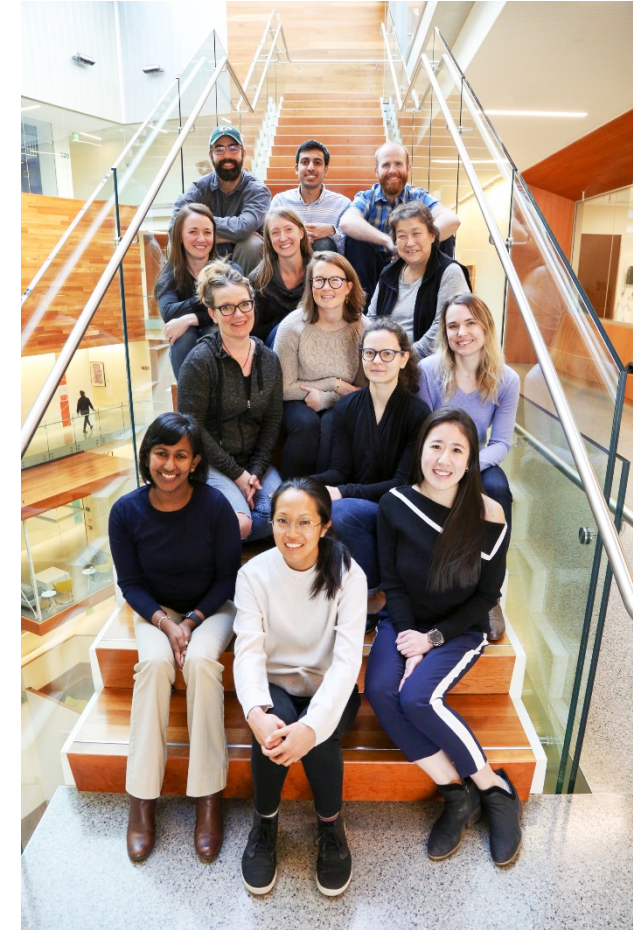
- Endogenous FP-tagging with CRISPR/Cas9 (38 lines)
 - Extensive QC for cell biological, genomic, and stem cell criteria
- Sharing cell lines, plasmids, and methods
 - Cell lines and plasmids - through Coriell and Addgene
 - Distribution to stem cell cores at major institutes
 - Methods and tutorials - MBoC and Stem cell Reports (Roberts et al., 2017, 2019)
 - Allencell.org



The Allen Institute for Cell Science team



The Stem Cell and Gene Editing group



Allen Institute for Cell Science: Gene Editing, Part I: Gene editing design

Amanda Haupt & Tanya Grancharova



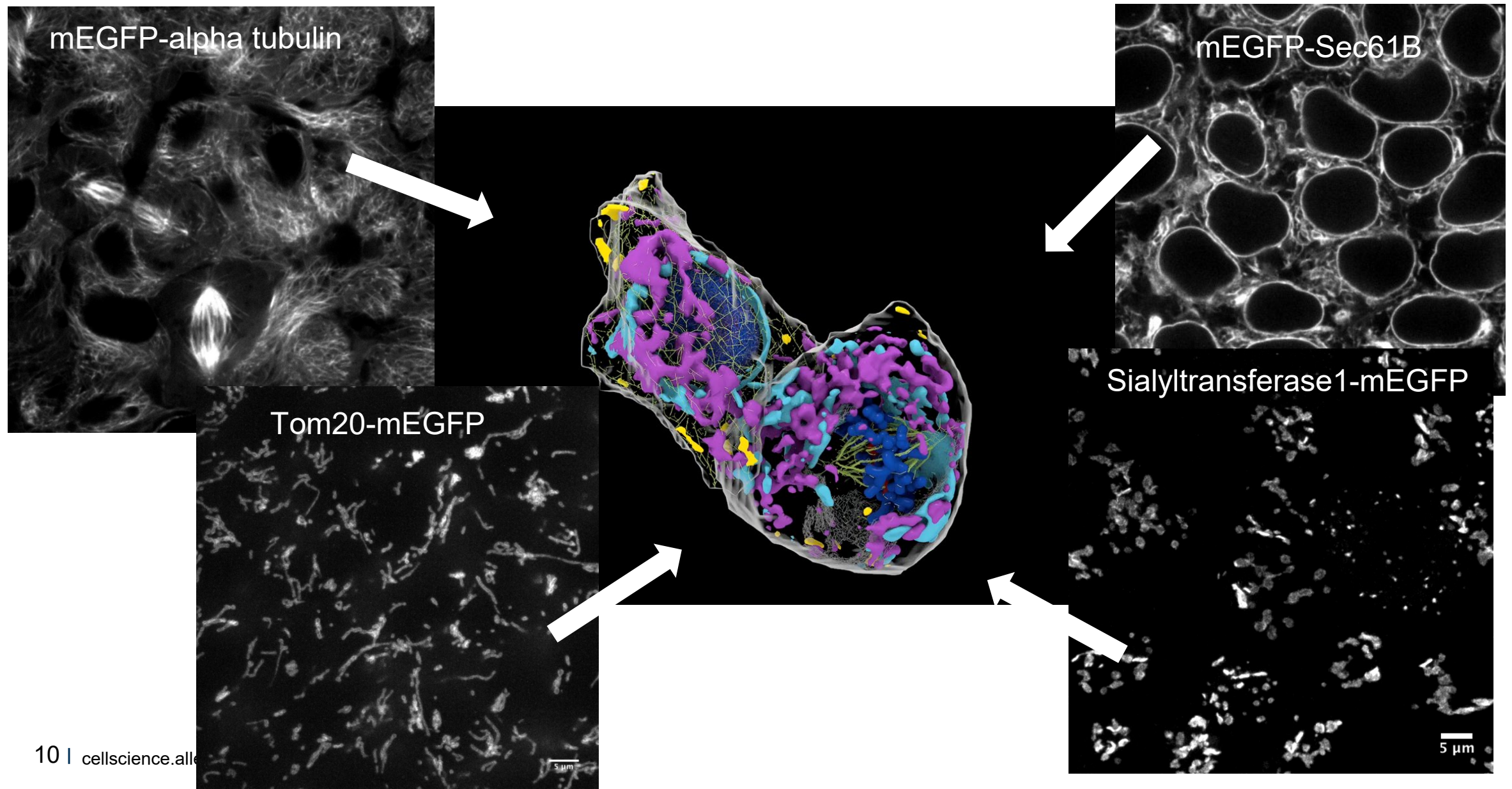
ALLEN INSTITUTE *for*
CELL SCIENCE

Outline

- Intro to the Allen Institute for Cell Science
- **Gene editing design**
 - Introduction
 - crRNA design
 - Donor template design
- Workflows and methods for generating high-quality clonal hiPS cell lines
- In-lab demonstrations
- Roundtable discussion



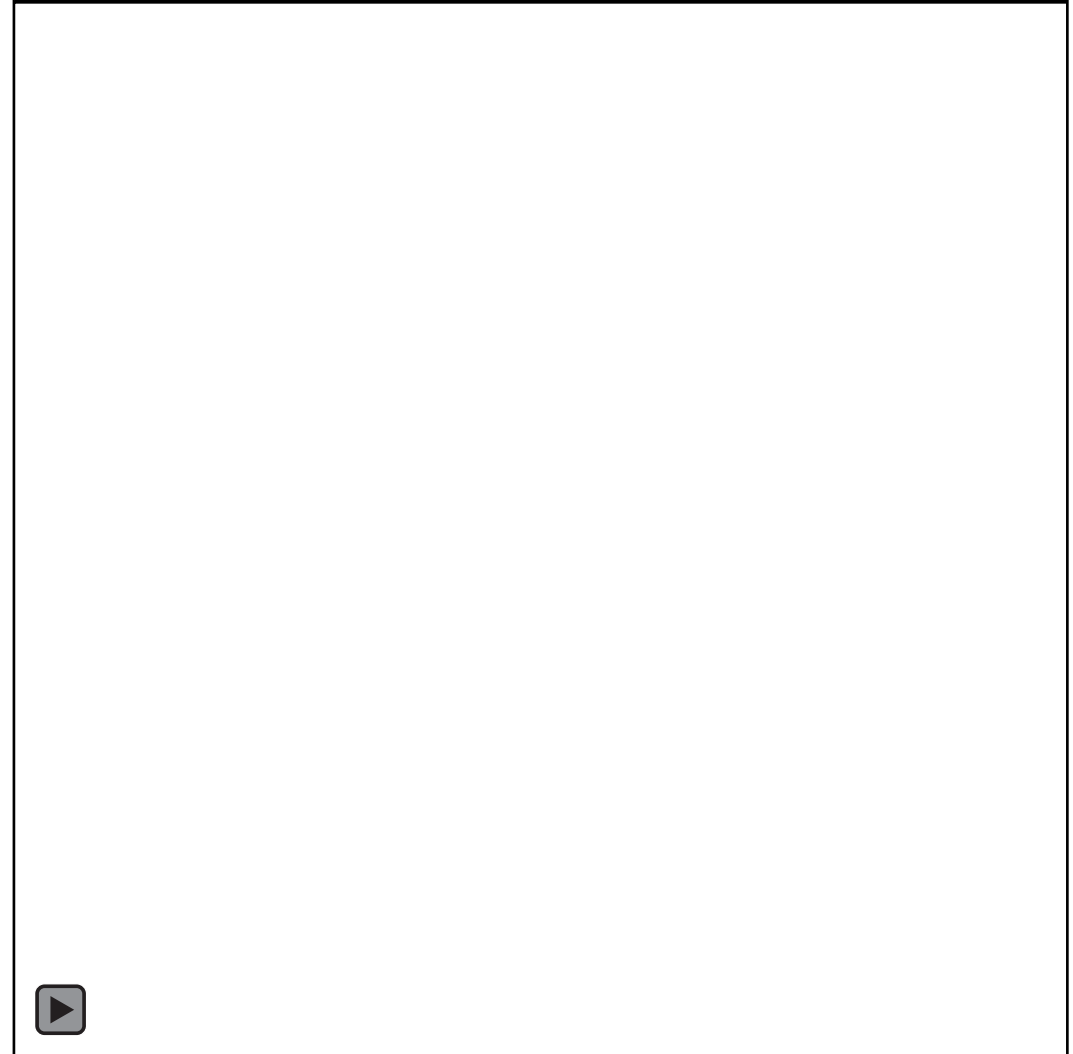
Allen Institute for Cell Science: Creating a collection of FP tagged hiPSC lines representing major structures of the cell



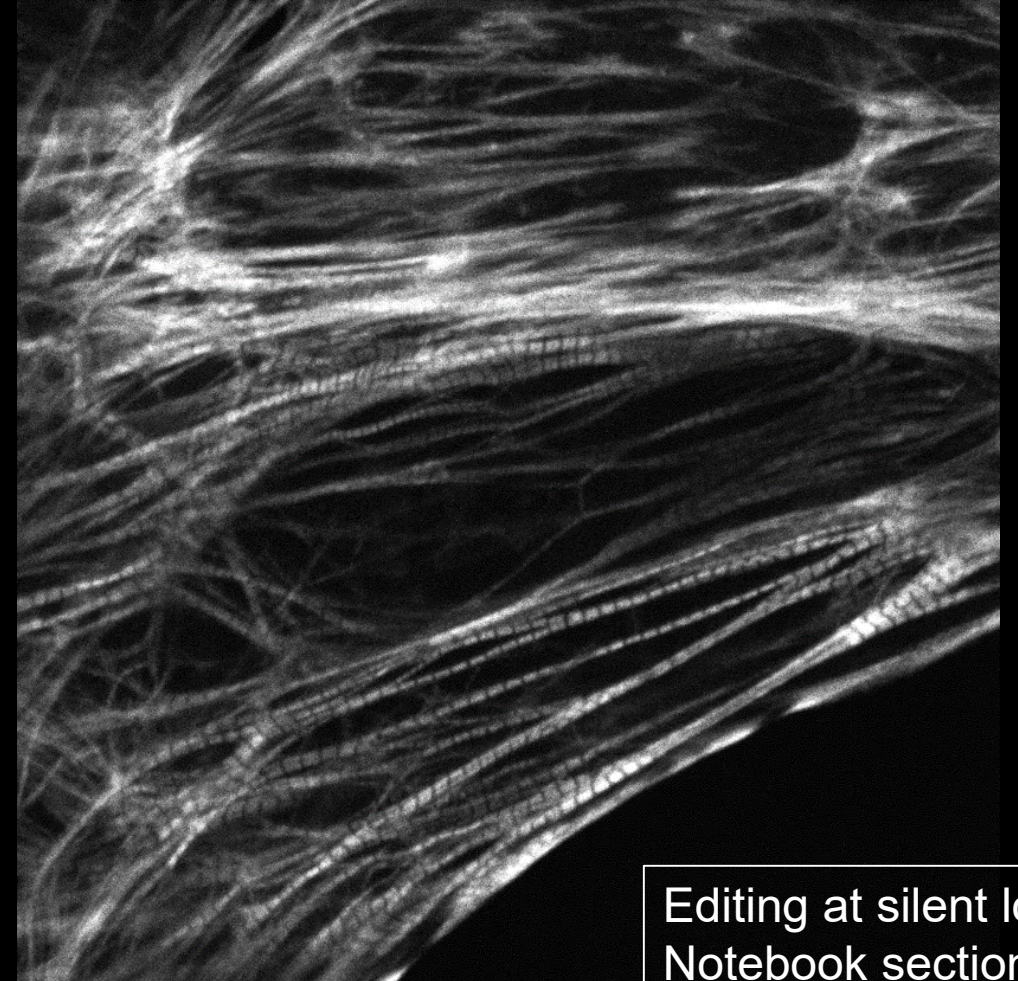
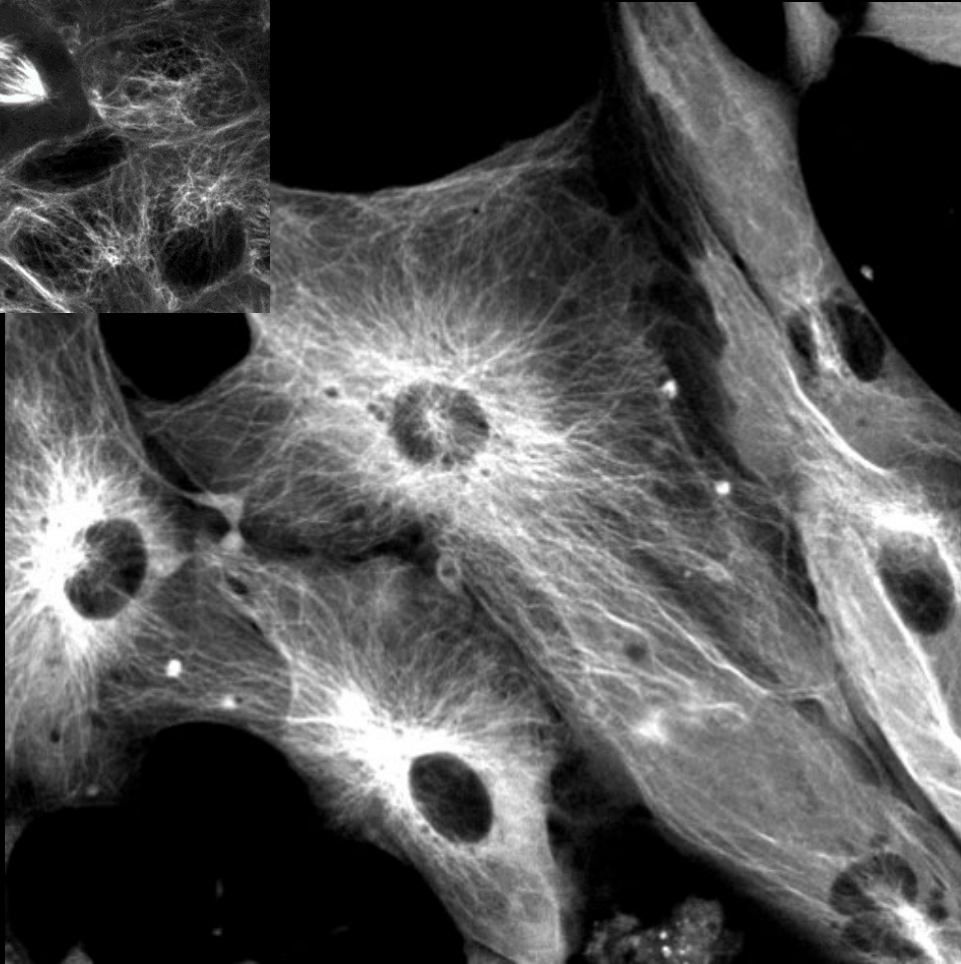
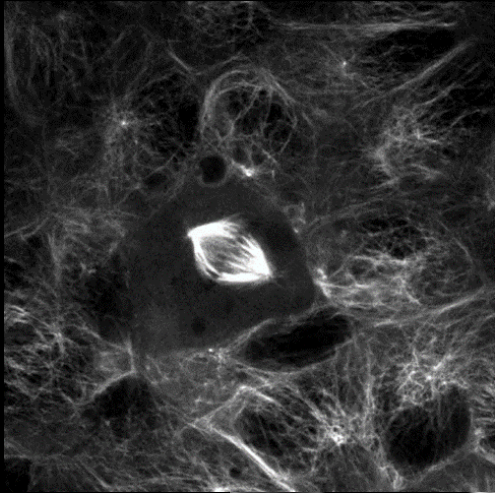
Endogenously tagged clonal cell lines generated

Structures Illuminated

- Nucleus (LMNB1)
- Nucleolus (FBL, NPM1, UBTF)
- Nuclear pores (NUP153)
- Chromatin (SMC1A, HIST1H2BJ)
- Mitochondria (TOMM20)
- Microtubules (TUBA1B)
- Centrosomes (CETN2)
- Endoplasmic Reticulum (SEC61B)
- Golgi (ST6GAL1)
- Lysosomes (LAMP1)
- Peroxisomes (SLC25A17)
- Autophagosomes (MAP1LC3B)
- Endosomes (RAB5A)
- Actins, myosins & filaments (ACTB, ACTN1, ACTN2, MYH10, VIM)
- Cell-cell junctions (TJP1/ZO1, DSP)
- Gap junctions (GJA1)
- Adherens junctions (CTNNB1)
- Matrix Adhesions (PXN)
- Stress Granules and P-Bodies (G3BP1, DCP1A)
- Transcription Factors (SOX2, POU5F1)
- Sub-nuclear structures (CBX1, CTCF, SMC1A)
- Cytoplasm
- Plasma Membrane
- Sarcomere (TTN, MYL7, MYL2, TNNT1, ACTN2)
- Sarcoplasmic reticulum (ATP2A2)



Harnessing the power of hiPSC



Editing at silent loci:
Notebook section 1C

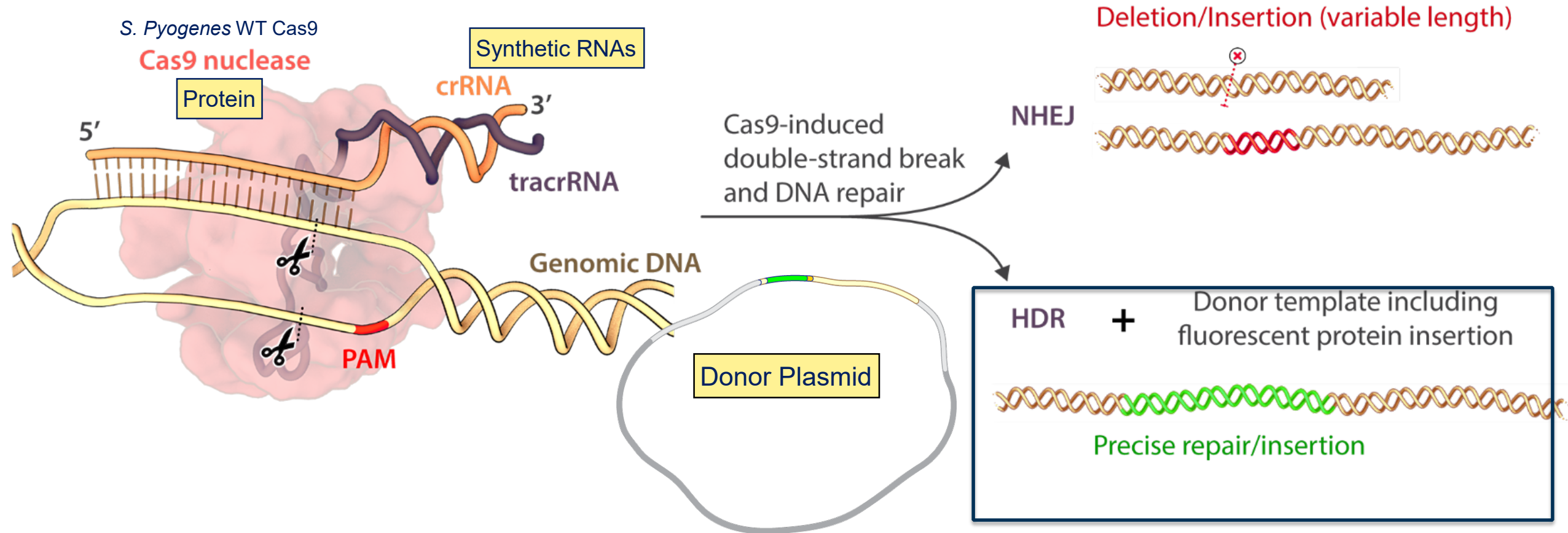
Cardiomyocytes from mEGFP-Alpha tubulin cell line

Cardiomyocytes from Troponin 1-mEGFP cell line

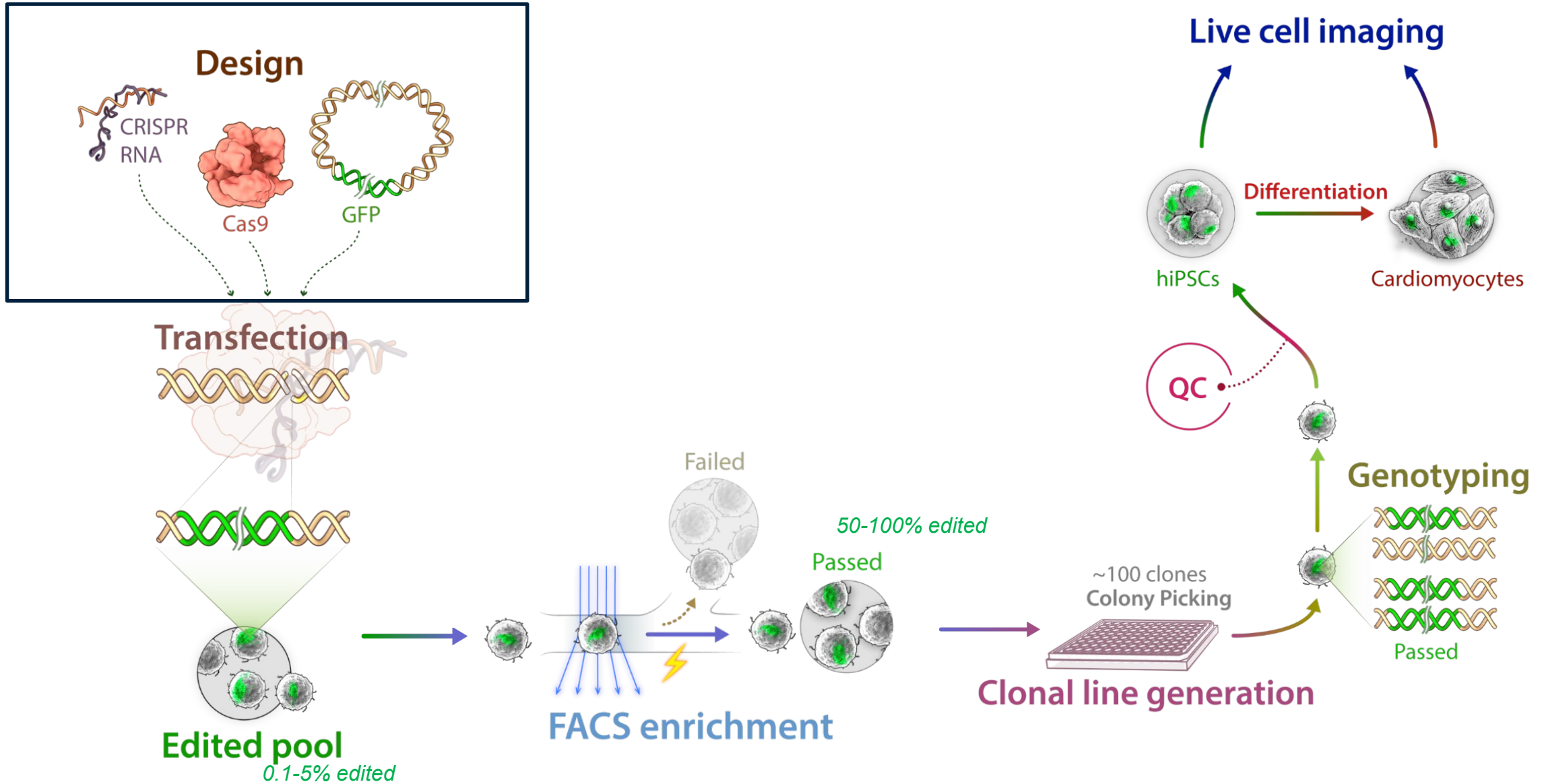
Cardiomyocyte movies: 100 ms intervals, 25s total 2D timelapse ~ real time - - Thank you Assay Dev Team!



CRISPR/Cas9 using the RNP Method: A gentle and effective way to introduce specific genomic insertions in stem cells



The Allen Institute for Cell Science Gene Editing Workflow



Outline

- Intro to the Allen Institute for Cell Science
- Gene editing design
 - Introduction
 - **crRNA design**
 - Donor template design
- Workflows and methods for generating high-quality clonal hiPS cell lines
- In-lab demonstrations
- Roundtable discussion



Editing design prerequisites

- **Select target gene, linker, and tag**



Editing design: Selecting a linker

- **Survey literature**
- **Review of linker design: Chen et al. 2013 (Section 1b in workbook)**
- **Short flexible linker: GTSGGS**

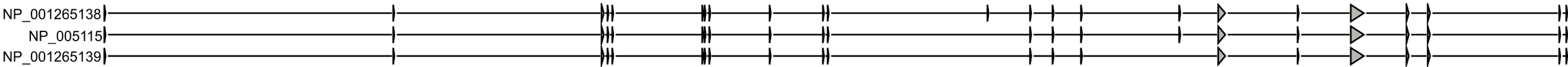


Editing design: Selecting a target gene, linker and tag

Gene: NUP153 (Nucleoporin Nup153)

Linker: YSDLE

Tag: mEGFP



Structure: Nuclear pores



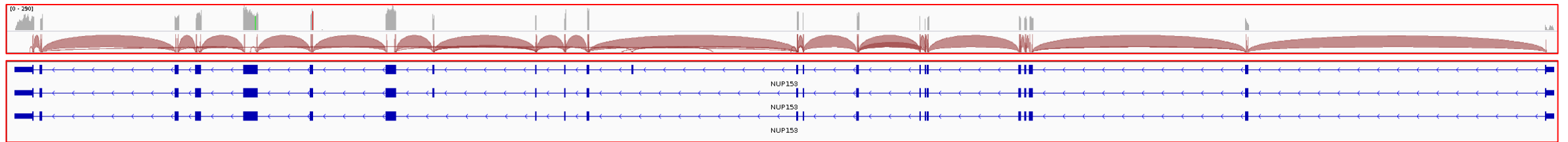
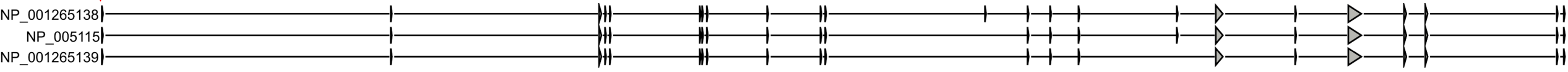
Editing design prerequisites

- Select target gene, linker, and tag
- **Acquire cell line specific data (recommended)**



Tag here

Editing design: Do you have isoform level information?



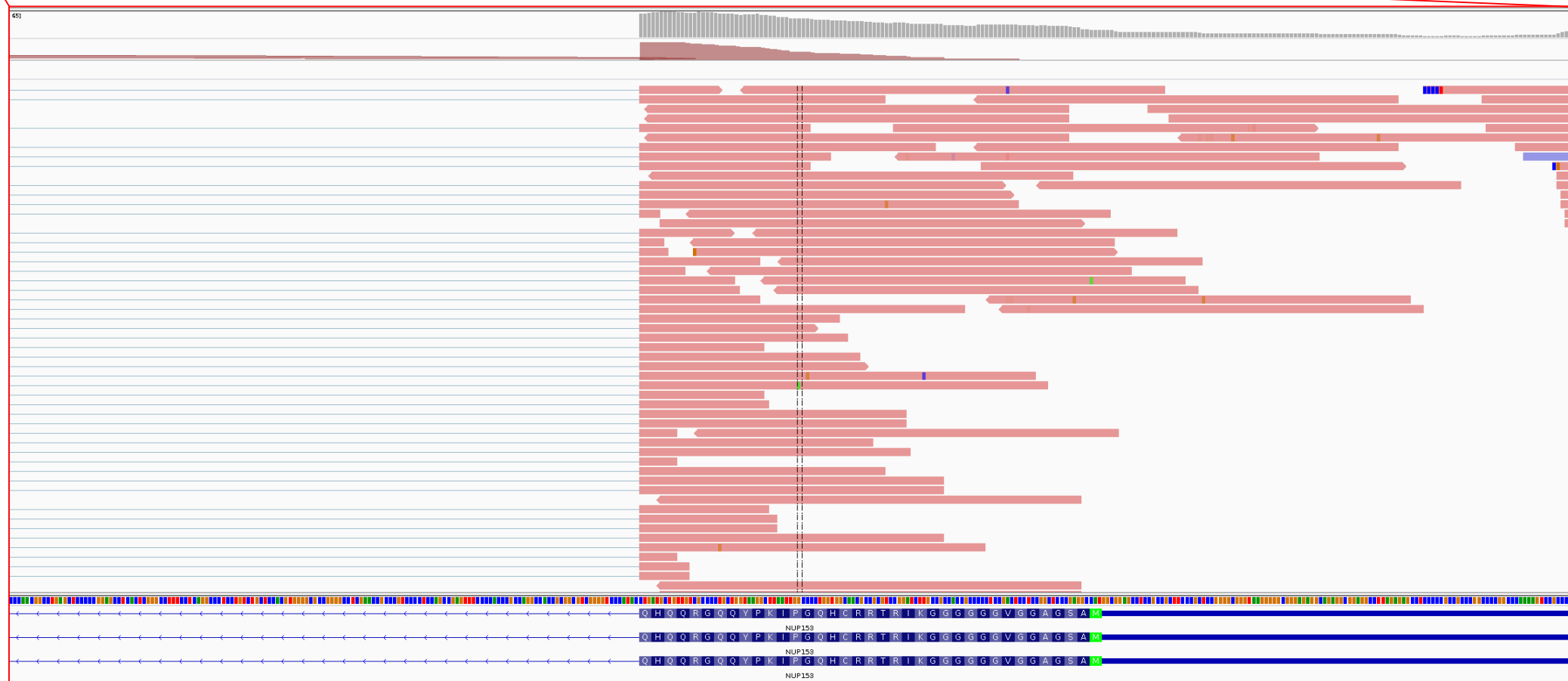
Tag here

Multiple isoforms w/ same N-terminus



Tag here

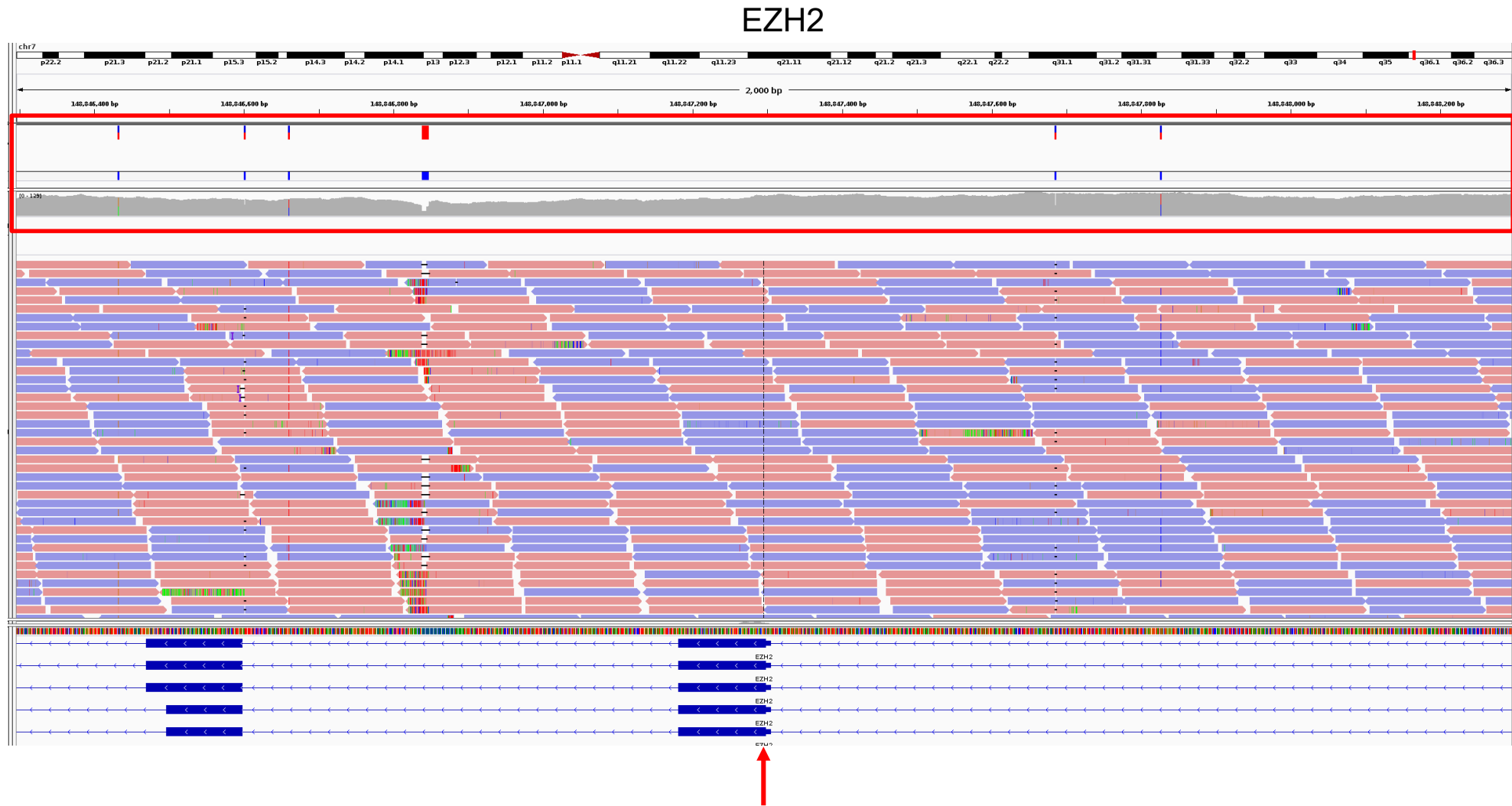
Editing design: Do you have isoform level information?



Tag here



Editing design: Do you have cell line variant information?



Editing design prerequisites

- Select target gene, linker, and tag
- Acquire cell line specific data (recommended)
- **Sequences and tools**
 - **Software to view/annotate/manipulate sequences**
 - Reference sequence, tag sequences, and plasmid backbone
 - crRNA design tool



Editing design prerequisites

- Select target gene, linker, and tag
- Acquire cell line specific data (recommended)
- **Sequences and tools**
 - Software to view/annotate/manipulate sequences
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Editing design prerequisites

- Select target gene, linker, and tag
- Acquire cell line specific data (recommended)
- **Sequences and tools**
 - Software to view/annotate/manipulate sequences
 - Reference sequence, tag sequences, and plasmid backbone
 - **crRNA design tool**



Editing design: Download sequences

- Download **NUP153 sequence** from NCBI <https://www.ncbi.nlm.nih.gov/gene/> in genbank format



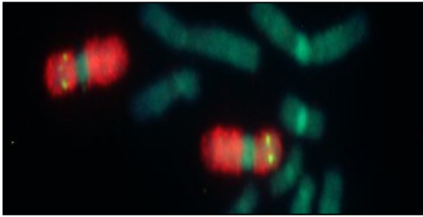
Download NUP153 sequence

Home - Gene - NCBI

https://www.ncbi.nlm.nih.gov/gene/

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- [Gene News](#)
- [Factsheet](#)

Representative queries

Find genes by...

free text

chromosome and symbol

Search text

human muscular dystrophy

(11[chr] OR 2[chr]) AND adh*[sym]

Homo sapiens NUP153

Homo sapiens nuclear pore complex protein Nup153

Drosophila melanogaster Nup153

Rattus norvegicus Nup153

Danio rerio nup153

Mus musculus Nup153

Bos taurus NUP153

Pongo abelii NUP153

Rattus norvegicus nuclear pore complex protein Nup153

Danio rerio nuclear pore complex protein Nup153

Mus musculus nuclear pore complex protein Nup153

Bos taurus nuclear pore complex protein Nup153

Pongo abelii nuclear pore complex protein Nup153

Sus scrofa NUP153

Pan troglodytes NUP153

Equus caballus NUP153

Macaca mulatta NUP153

Dasyatis novemcinctus NUP153

Oryctolagus cuniculus NUP153

Sus scrofa nuclear pore complex protein Nup153



Download NUP153 sequence

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[https://www.ncbi.nlm.nih.gov/gene/?term=Homo+sapiens+NUP153](#)
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GENE
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[NUP153 – nucleoporin 153](#)

[Homo sapiens \(human\)](#)

Also known as: HNUP153, N153

GeneID: 9972

[RefSeq transcripts \(8\)](#)
[RefSeq proteins \(8\)](#)
[PubMed \(146\)](#)

[RefSeq transcripts](#)
[+](#)

[RefSeq proteins](#)
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[Caenorhabditis remanei \(1\)](#)
[\[Candida\] auris \(1\)](#)
[\[Candida\] pseudohaemulonius \(1\)](#)

Find related data

Database:

Search details

```
((("Homo sapiens"[Organism] OR Homo sapiens[All Fields]) AND NUP153[All Fields]) AND alive[prop])
```

[See more...](#)

Search results

Items: 1 to 20 of 132

[https://www.ncbi.nlm.nih.gov/gene/9972](#)

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Page of 7
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[Last >>](#)



Download NUP153 sequence

The screenshot shows the NCBI Gene database page for NUP153 nucleoporin 153 [Homo sapiens (human)]. The browser address bar shows the URL: <https://www.ncbi.nlm.nih.gov/gene/9972>. The page header includes the NCBI logo, navigation links (Resources, How To), and a sign-in link. The main content area displays the gene name, ID (9972), and update date (13-Feb-2019). A sidebar on the right contains a table of contents with links to various sections. The main content area is divided into two sections: a summary and a detailed description.

Gene [Advanced](#) [Help](#)

[Full Report](#) [Send to:](#) [Hide sidebar >>](#)

NUP153 nucleoporin 153 [*Homo sapiens* (human)]

Gene ID: 9972, updated on 13-Feb-2019

Summary

Official Symbol NUP153 provided by [HGNC](#)
Official Full Name nucleoporin 153 provided by [HGNC](#)
Primary source [HGNC:HGNC:8062](#)
See related [Ensembl:ENSG00000124789](#) [MIM:603948](#)
Gene type protein coding
RefSeq status REVIEWED
Organism [Homo sapiens](#)
Lineage Eukaryota; Metazoa; Chordata; Craniata; Vertebrata; Euteleostomi; Mammalia; Eutheria; Euarchontoglires; Primates; Haplorrhini; Catarrhini; Hominidae; Homo
Also known as N153; HNUP153
Summary Nuclear pore complexes regulate the transport of macromolecules between the nucleus and cytoplasm. They are composed of at least 100 different polypeptide subunits, many of which belong to the nucleoporin family. Nucleoporins are glycoproteins found in nuclear pores and contain characteristic pentapeptide XFXFG repeats as well as O-linked N-acetylglucosamine residues oriented towards the cytoplasm. The protein encoded by this gene has three distinct domains: a N-terminal region containing a pore targeting and an RNA-binding domain domain, a central region containing multiple zinc finger motifs, and a C-terminal region containing multiple XFXFG repeats. Alternative splicing results in multiple transcript variants of this gene. [provided by RefSeq, May 2013]
Expression Ubiquitous expression in testis (RPKM 19.5), bone marrow (RPKM 18.1) and 25 other tissues [See more](#)
Orthologs [mouse](#) [all](#)

Table of contents

- Summary
- Genomic context
- Genomic regions, transcripts, and products
- Expression
- Bibliography
- Phenotypes
- Variation
- HIV-1 interactions
- Pathways from BioSystems
- Interactions
- General gene information
 - Markers, Homology, Gene Ontology
- General protein information
- NCBI Reference Sequences (RefSeq)
- Related sequences
- Additional links



Download NUP153 sequence

<https://www.ncbi.nlm.nih.gov/gene/9972>

Annotation release	Status	Assembly	Chr	Location
109	current	GRCh38.p12 (GCF_000001405.38)	6	NC_000006.12 (17615035..17706834, complement)
105	previous assembly	GRCh37.p13 (GCF_000001405.25)	6	NC_000006.11 (17615266..17707065, complement)

Chromosome 6 - NC_000006.12

Genomic regions, transcripts, and products

Genomic Sequence: [NC_000006.12](#) Chromosome 6 Reference GRCh38.p12 Primary Assembly

Go to nucleotide: [Graphics](#) [FASTA](#) [GenBank](#)

NC_000006.12 | Find:

Genes, NCBI Homo sapiens Annotation Release 109, 2018-03-27

NUP153

NM_001278210.1
 NM_005124.3
 NM_001278209.1
 XM_017011594.2
 XM_005249507.3
 XM_006715290.3
 XM_011515028.3
 XM_006715291.4

LOC105374952
 NR_134616.1
 NR_134618.1

NP_001265139.1
 NP_005115.2
 NP_001265138.1
 XP_016867083.1
 XP_005249564.1
 XP_006715353.1
 XP_011513330.1
 XP_006715354.1

RNU6-190P
 XR_002956328.1
 FAM8A1
 NP_057339.1

Biological regions, aggregate, NCBI NCBI Homo sapiens Annotation Release 109, 2018-03-27
 Warning: No track data found in this range
 Genes, Ensembl release 95

1000 Genomes Browser (GRCh37.p13)
 Ensembl
 UCSC

Related information

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 BioAssay by Target (Summary)
 BioAssay, by Gene target
 BioAssays, RNAi Target, Tested
 BioProjects
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 ClinVar
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 Full text in PMC
 Full text in PMC_nucleotide
 Functional Class
 Gene neighbors
 Genes with a similar H3K4me3 profile
 Genome



Download NUP153 sequence

Homo sapiens chromosome 6, X

https://www.ncbi.nlm.nih.gov/nucore/NC_000006.12?report=genbank&from

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Nucleotide Nucleotide Search

Advanced Help

The Nucleotide database will include EST and GSS sequences in early 2019. [Read more.](#)

GenBank Send to

Homo sapiens chromosome 6, GRCh38.p12 Primary Assembly

NCBI Reference Sequence: NC_000006.12

[FASTA](#) [Graphics](#)

LOCUS NC_000006 91800 bp DNA linear CON 26-MAR-2018

DEFINITION Homo sapiens chromosome 6, GRCh38.p12 Primary Assembly.

ACCESSION NC_000006 REGION: complement(17615035..17706834)

VERSION NC_000006.12

DBLINK BioProject: [PRJNA168](#)
Assembly: [GCF_000001405.38](#)

KEYWORDS RefSeq.

SOURCE Homo sapiens (human)

ORGANISM [Homo sapiens](#)
Eukaryota; Metazoa; Chordata; Craniata; Vertebrata; Euteleostomi;
Mammalia; Eutheria; Euarchontoglires; Primates; Haplorrhini;
Catarrhini; Hominidae; Homo.

REFERENCE 1 (bases 1 to 91800)
CONSTRM International Human Genome Sequencing Consortium
TITLE Finishing the euchromatic sequence of the human genome
JOURNAL Nature 431 (7011), 931-945 (2004)
PUBMED [15496913](#)

REFERENCE 2 (bases 1 to 91800)
AUTHORS Mungall,A.J., Palmer,S.A., Sims,S.K., Edwards,C.A., Ashurst,J.L.,
Wilming,L., Jones,M.C., Horton,R., Hunt,S.E., Scott,C.E.,
Gilbert,J.G., Clamp,M.E., Bethel,G., Milne,S., Ainscough,R.,

Change region shown

☐ Whole sequence (abbreviated view)
☒ Selected region
from: 17615035 to: 17706834
[Update View](#)

Customize view

☐ Abbreviated view
☒ Customize

Basic Features

☐ Default features
☒ Gene, RNA, and CDS features only

Features added by NCBI

☐ 1 SNP

Display options

☒ Show sequence
☒ Show reverse complement
[Update View](#)

Analyze this sequence



Download NUP153 sequence

Homo sapiens chromosome 6, X

https://www.ncbi.nlm.nih.gov/nucore/NC_000006.12?report=genbank&from

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Nucleotide Nucleotide Search

Advanced Help

The Nucleotide database will include EST and GSS sequences in early 2019. [Read more.](#)

GenBank

Showing 95.80kb region from base 17613035 to 17708834.

Homo sapiens chromosome 6, GRCh38.p12 Primary Assembly

NCBI Reference Sequence: NC_000006.12

[FASTA](#) [Graphics](#)

LOCUS NC_000006 95800 bp DNA linear CON 26-MAR-2018

DEFINITION Homo sapiens chromosome 6, GRCh38.p12 Primary Assembly.

ACCESSION [NC_000006](#) REGION: complement(17613035..17708834)

VERSION NC_000006.12

DBLINK BioProject: [PRJNA168](#)

Assembly: [GCF_000001405.38](#)

KEYWORDS RefSeq.

SOURCE Homo sapiens (human)

ORGANISM [Homo sapiens](#)

Eukaryota; Metazoa; Chordata; Craniata; Vertebrata; Euteleostomi; Mammalia; Eutheria; Euarchontoglires; Primates; Haplorrhini; Catarrhini; Hominidae; Homo.

REFERENCE 1 (bases 1 to 95800)

CONSTRM International Human Genome Sequencing Consortium

TITLE Finishing the euchromatic sequence of the human genome

JOURNAL Nature 431 (7011), 931-945 (2004)

PUBMED [15496913](#)

REFERENCE 2 (bases 1 to 95800)

AUTHORS Mungall,A.J., Palmer,S.A., Sims,S.K., Edwards,C.A., Ashurst,J.L.,

Change region shown

☐ Whole sequence (abbreviated view)

☒ Selected region

from: 17613035 to: 17708834

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Customize view

☐ Abbreviated view

☒ Customize

Basic Features

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Features added by NCBI

☐ 1 SNP

Display options

☒ Show sequence

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Analyze this sequence



Download NUP153 sequence

[illegible]

Download NUP153 sequence (Ensembl)

The screenshot displays the Ensembl genome browser interface for the NUP153 gene (ENSG00000124789) on human chromosome 6. The browser window has a dark blue header with the Ensembl logo and the gene name. The left sidebar contains a navigation menu with categories like Ontologies, Genetic Variation, and Gene expression. The main content area is titled 'Summary' and provides key information about the gene, including its name, CCDS set, UniProtKB identifiers, RefSeq ID, Ensembl version, and other assemblies. Below the summary, there is a section for 'Gene type' and 'Annotation method'. A blue banner below the summary encourages users to 'Go to Region in Detail for more tracks and navigation options (e.g. zooming)'. The bottom section shows a genomic track view with a scale from 17.62Mb to 17.70Mb. It includes tracks for Genes (FAM8A1-201, RNU6-190P-201, AL138824.19, AL157776.15, AL138724.12), Contigs, and Regulatory Build. The NUP153 gene is highlighted in green, showing its protein-coding structure with exons and introns. The forward strand is indicated at the top right of the track view.

Summary

Name [NUP153](#) (HGNC Symbol)

CCDS This gene is a member of the Human CCDS set: [CCDS4541.1](#), [CCDS64359.1](#), [CCDS75407.1](#)

UniProtKB This gene has proteins that correspond to the following UniProtKB identifiers: [P49790](#)

RefSeq Overlapping RefSeq Gene ID [9972](#) matches and has similar biotype of protein_coding

Ensembl version ENSG00000124789.11

Other assemblies This gene maps to [17,615,266-17,707,065](#) in GRCh37 coordinates.
View this locus in the GRCh37 archive: [ENSG00000124789](#)

Gene type Protein coding

Annotation method Annotation for this gene includes both automatic annotation from Ensembl and [Havana](#) manual curation, see [article](#).

Go to Region in Detail for more tracks and navigation options (e.g. zooming)

Genomic Track View:

- Scale: 17.62Mb to 17.70Mb (111.80 kb)
- Forward strand
- Genes (Comprehensive set...): FAM8A1-201 > protein coding, RNU6-190P-201 > snRNA, AL138824.19 >, AL157776.15 >, AL138724.12 > antisense
- Contigs (Comprehensive set...): AL138824.19 >, AL157776.15 >, AL138724.12 >
- Regulatory Build: Shows various regulatory elements (yellow and red blocks)

https://uswest.ensembl.org/Homo_sapiens/View/Configure/Gene?db=core;g=ENSG00000124789;r=6:17615035-17706834



Download NUP153 sequence (Ensembl)

The screenshot shows the Ensembl genome browser interface for the NUP153 gene (ENSG00000124789) on human chromosome 6. The 'Export data' dialog is open, displaying the 'Export Configuration - Feature List'. The 'Gene to export' field is set to 'ENSG00000124789 (NUP153)'. The 'Output' dropdown menu is open, showing options: 'FASTA sequence' (selected), 'Bed Format', 'FASTA sequence', 'Feature File' (with sub-options: CSV, Tab, GTF, GFF, GFF3), 'Flat File' (with sub-options: EMBL, GenBank), and 'PIP (%age identity plot)' (with sub-options: Pipmaker / zPicture format, Vista Format). The 'Strand' field is empty. The '5' Flanking sequence (upstream):' and '3' Flanking sequence (downstream):' fields are empty. A note states 'Fields marked * are required'. The 'Options for FASTA sequence' section shows 'Genomic:' as the selected option. The background shows the gene track with various features and a 'Regulatory Build' track at the bottom.

Export Configuration - Feature List

Tip

For sequence export, please go to the relevant sequence page (see lefthand menu) and use the new "Download sequence" button

Gene to export: ENSG00000124789 (NUP153)

Output: FASTA sequence *

Strand:

5' Flanking sequence (upstream):

3' Flanking sequence (downstream):

Fields marked * are required

Options for FASTA sequence

Genomic:

Feature File

- CSV (Comma separated values)
- Tab separated values
- GTF (Gene Transfer Format)
- GFF (Generic Feature Format)
- GFF3 (Generic Feature Format Version 3)

Flat File

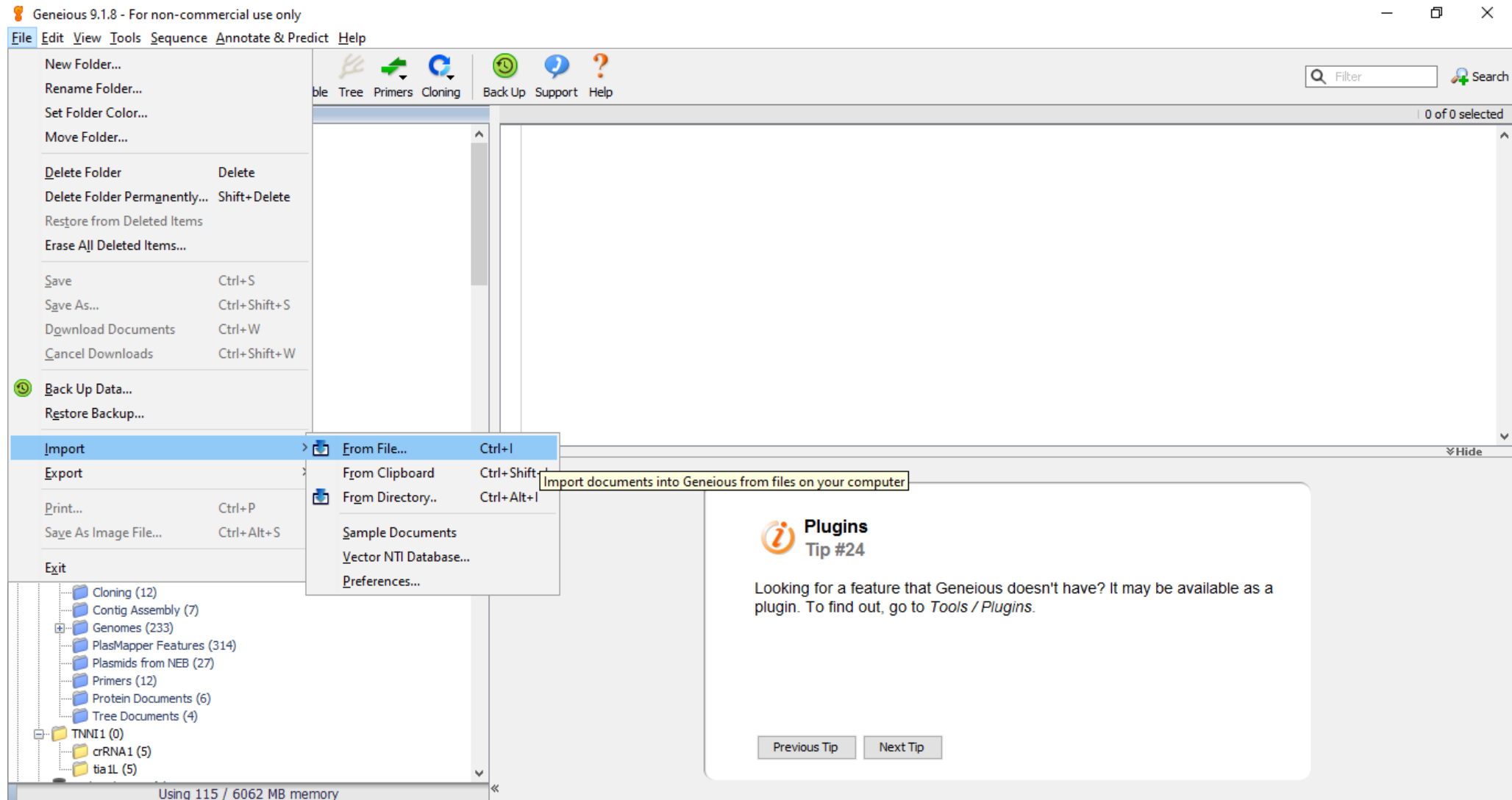
- EMBL
- GenBank

PIP (%age identity plot)

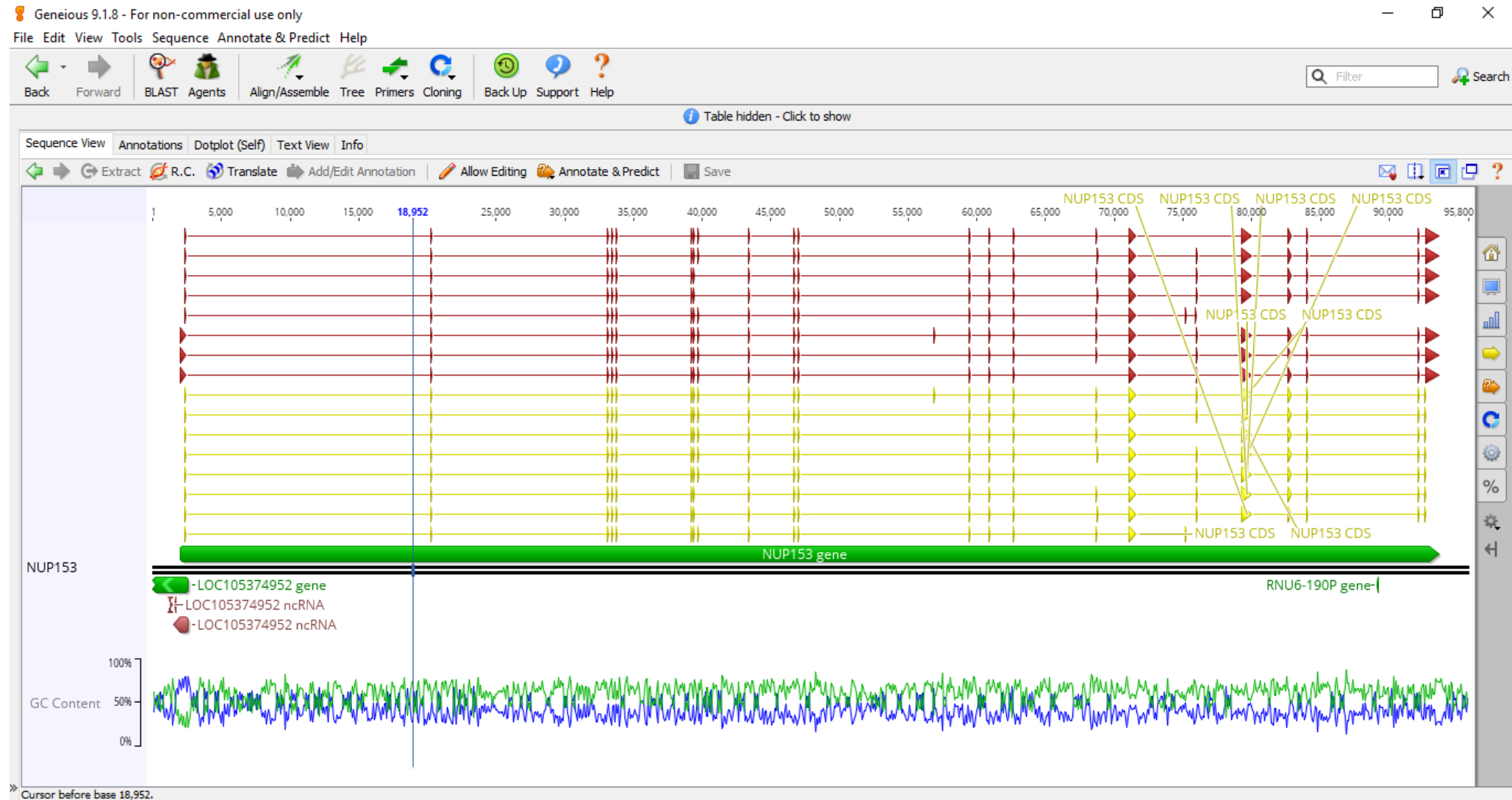
- Pipmaker / zPicture format
- Vista Format



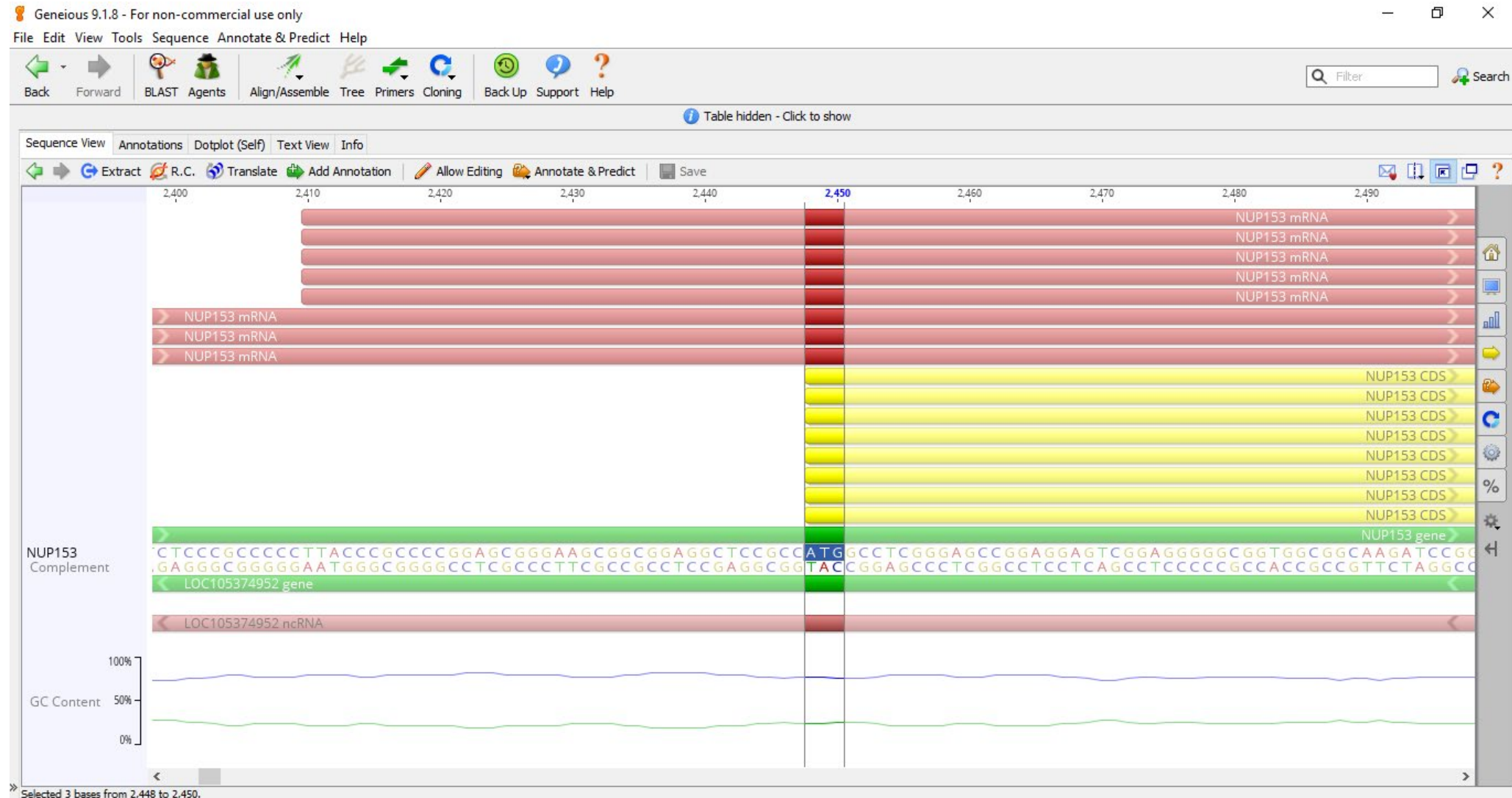
Editing design: Import sequence in Geneious



Editing design: Zoom to terminus you want to tag



Editing design: Zoom to terminus you want to tag



Editing design: Zoom to terminus you want to tag

Geneious 9.1.8 - For non-commercial use only

File Edit View Tools Sequence Annotate & Predict Help

Back Forward BLAST Agents Align/Assemble Tree Primers Cloning Back Up Support Help

Filter Search

Table hidden - Click to show

Sequence View Annotations Dotplot (Self) Text View Info

Extract R.C. Translate Add/Edit Annotation Allow Editing Annotate & Predict Save

2,400 2,410 2,420 2,430 2,440 2,450 2,460 2,467 2,470 2,480 2,490

NUP153 mRNA

NUP153 mRNA

NUP153 mRNA

NUP153 mRNA

NUP153 mRNA

NUP153 mRNA

NUP153 mRNA

NUP153 CDS

NUP153 CDS

NUP153 CDS

NUP153 CDS

NUP153 CDS

NUP153 CDS

NUP153 CDS

NUP153 CDS

NUP153 gene

Insertion b/w these two bases—

NUP153 Complement

LOC105374952 gene

LOC105374952 ncRNA

GC Content

100%

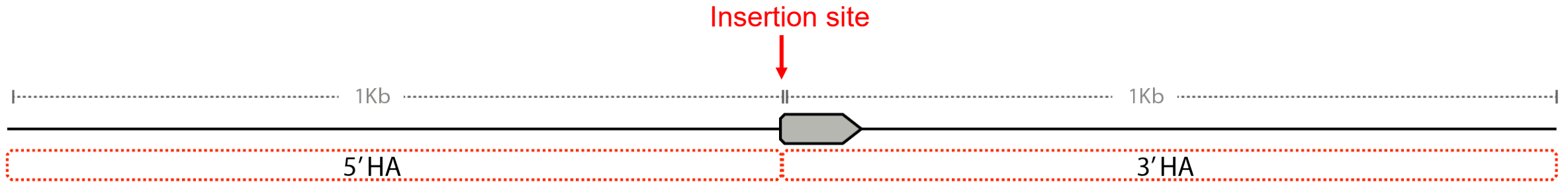
50%

0%

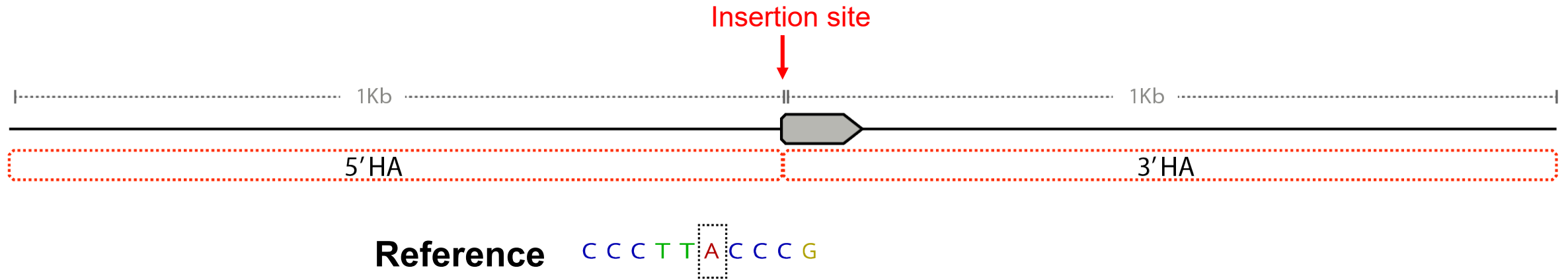
Cursor before base 2,467.



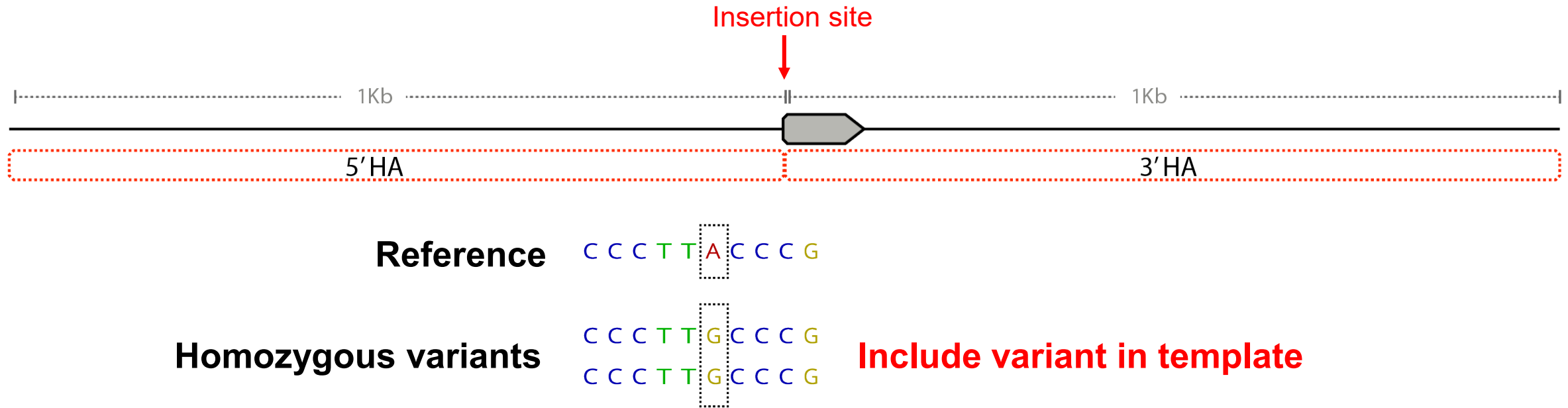
Editing design: Include cell line specific variants



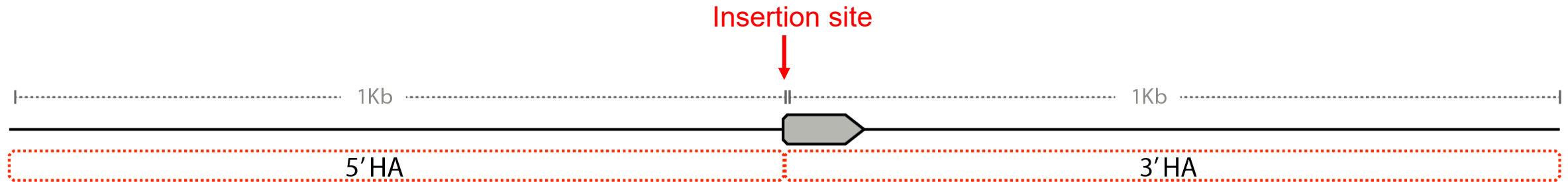
Editing design: Include cell line specific variants



Editing design: Include cell line specific variants



Editing design: Include cell line specific variants



Reference

C C C T T A C C C G

Homozygous variants

C C C T T G C C C G
C C C T T G C C C G

Include variant in template

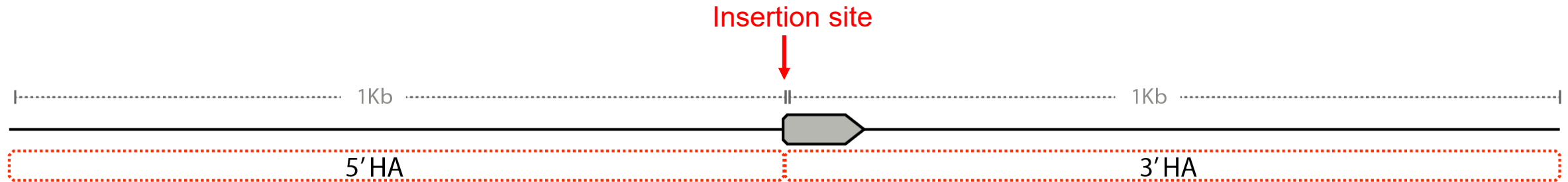
Heterozygous variants

C C C T T A C C C G
C C C T T G C C C G

Optional inclusion of variant in template



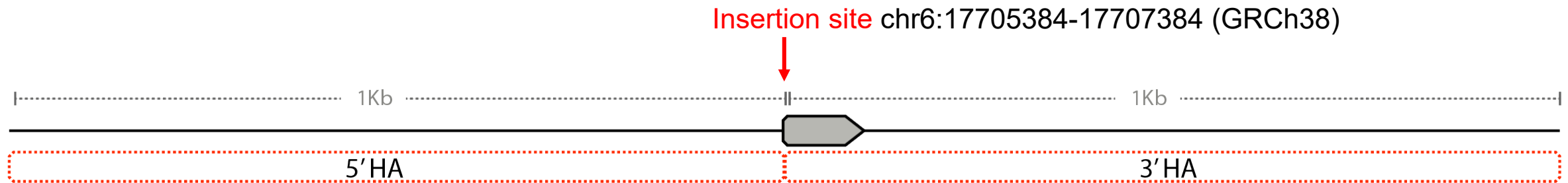
Editing design: Include cell line specific variants



Reference	C C C T T A C C C G	
Homozygous variants	C C C T T G C C C G C C C T T G C C C G	Include variant in template
Heterozygous variants	C C C T T A C C C G C C C T T G C C C G	Optional inclusion of variant in template
Heterozygous variants	C C C T T C C C C G C C C T T T C C C G	Select one variant to include in template



Editing design: Include cell line specific variants

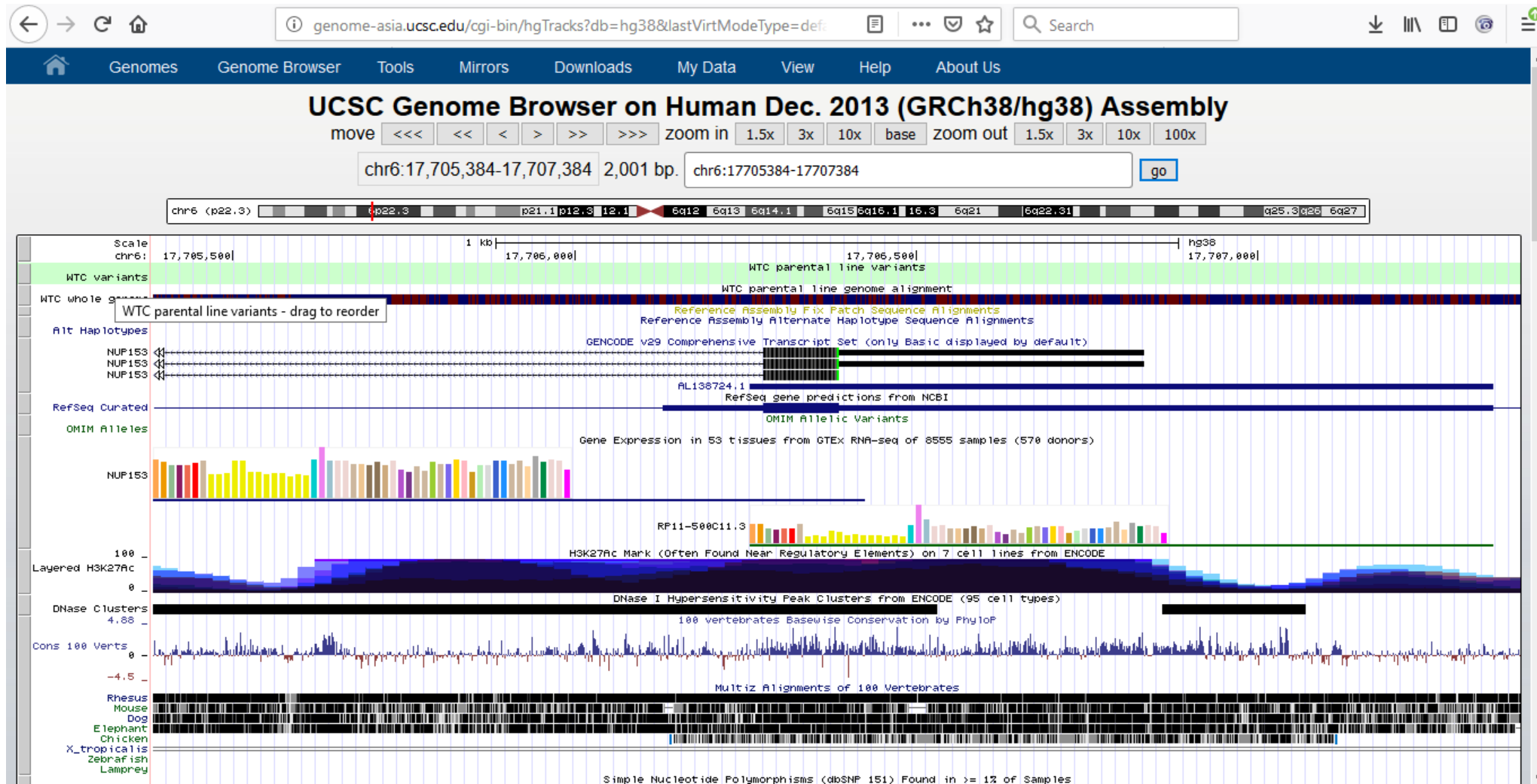


Reference	C C C T T A C C C G	
Homozygous variants	C C C T T G C C C G C C C T T G C C C G	Include variant in template
Heterozygous variants	C C C T T A C C C G C C C T T G C C C G	Optional inclusion of variant in template
Heterozygous variants	C C C T T C C C C G C C C T T T C C C G	Select one variant to include in template

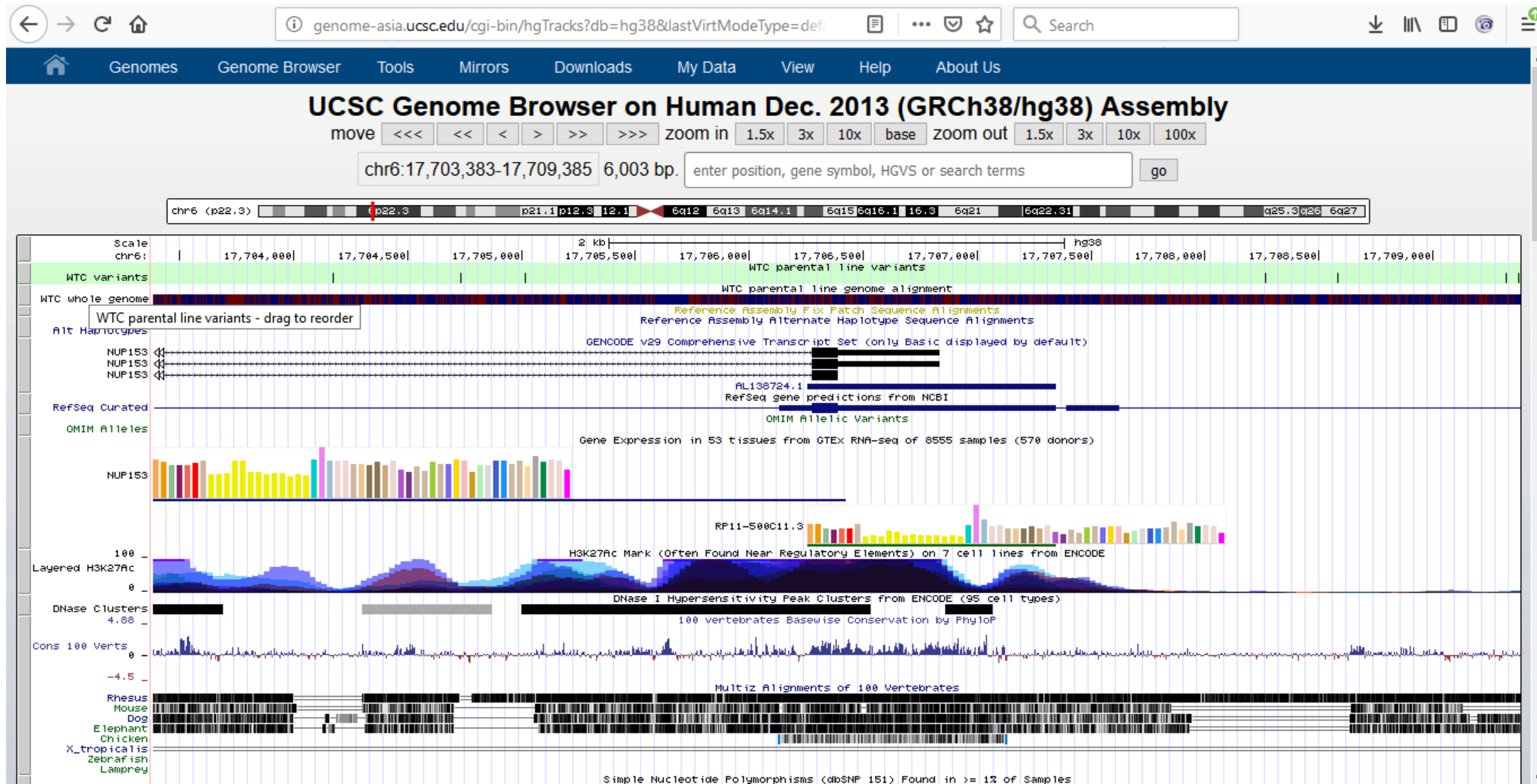
WTC-11 whole genome variants: <https://www.allencell.org/genomics.html>



Editing design: Include cell line specific variants

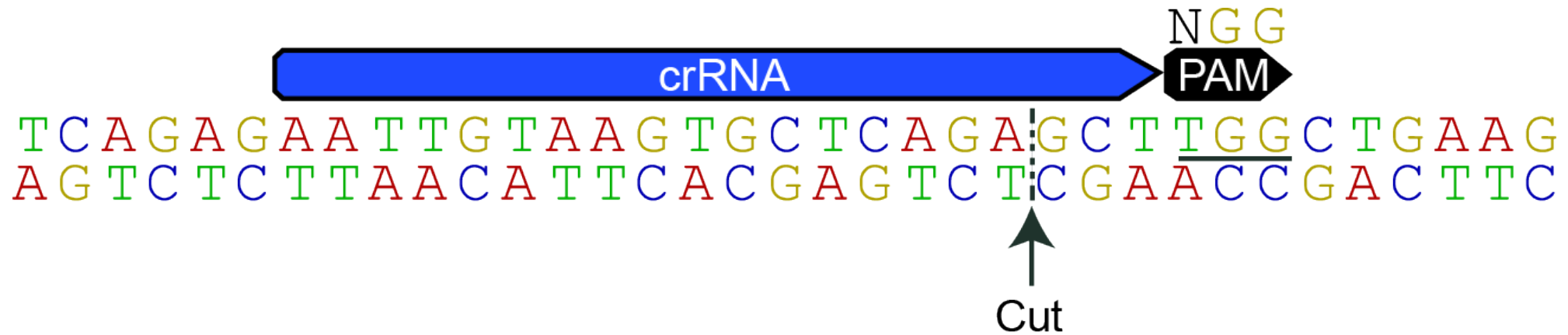


Editing design: Include cell line specific variants



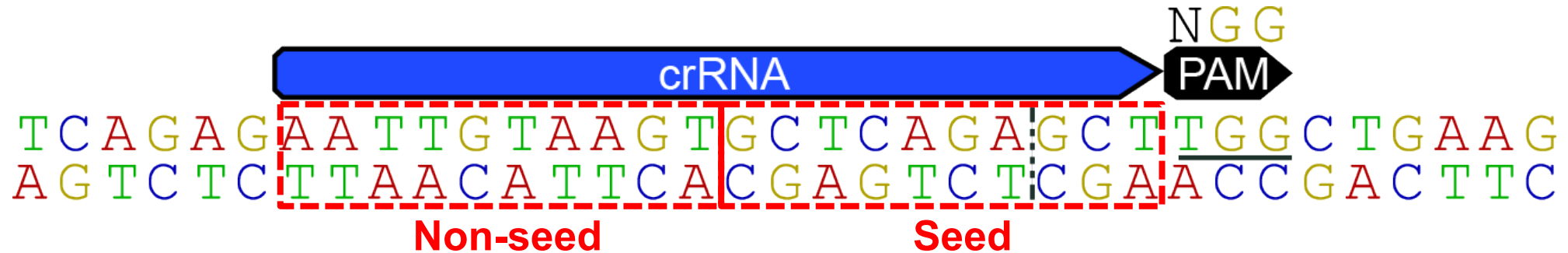
Editing design: Find CRISPR guides

Guide is 20nt sequence adjacent to NGG PAM (SpCas9)



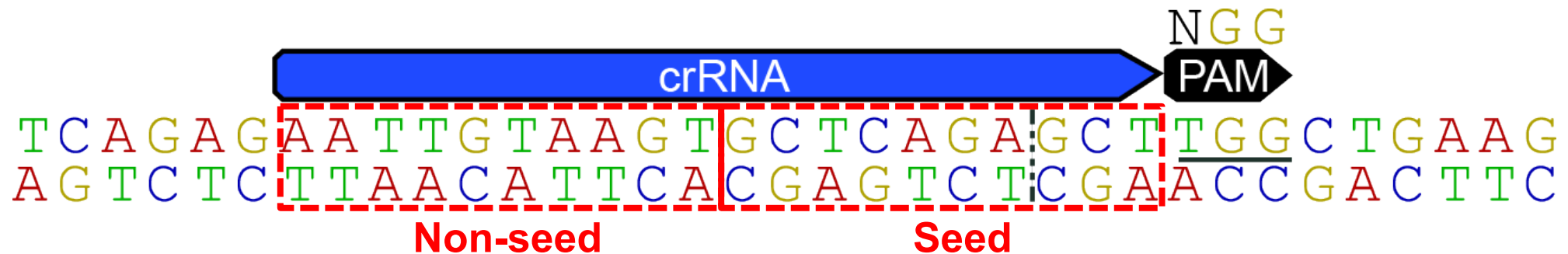
Editing design: Find CRISPR guides

Guide is 20nt sequence adjacent to NGG PAM (SpCas9)



Editing design: Selecting CRISPR guides

- Select guides by balancing:
 - Off-targets



Editing design: Selecting CRISPR guides (Avoiding off-targets)

		crRNA	PAM
Avoid	On-target	A A T T G T A A G T G C T C A G A G C T <u>T G G</u>	
	Off-target	A A T T G T A A G T G C T C A G A G C T T G G	
Avoid	On-target	A A T T G T A A G T G C T C A G A G C T <u>T G G</u>	
	Off-target	A A T T G T A A G T G C T C A G A G C T T [*] A G	

Avoid guides with perfect off-target matches (adjacent to NGG or NAG PAM)



Editing design: Selecting CRISPR guides (Avoiding off-targets)

Avoid	On-target	A A T T G T A A G T G C T C A G A G C T <u>T G G</u>
	Off-target	A A T T G T A A G T G C T C A G A G C T T G G
Avoid	On-target	A A T T G T A A G T G C T C A G A G C T <u>T G G</u>
	Off-target	A A T T G T A A G T G C T C A G A G C T T [*] A G

Avoid guides with perfect off-target matches (adjacent to NGG or NAG PAM)

	On-target	A A T T G T A A G T G C T C A G A G C T <u>T G G</u>
	Off-target	A A T [*] G G T T A G T G C T C A G A G C T T G G
Better	On-target	A A T T G T A A G T G C T C A G A G C T <u>T G G</u>
	Off-target	A A T T G T A A G T G C T C A G T G C C T G G

Prioritize guides that have no off-targets with perfectly matching seed regions



Editing design: Selecting CRISPR guides (Avoiding off-targets)

		<div>crRNA</div> <div>PAM</div>
Avoid	On-target	AA TTG TAAG TGC TCAG AGC T <u>TGG</u>
	Off-target	AA TTG TAAG TGC TCAG AGC C T TGG
Avoid	On-target	AA TTG TAAG TGC TCAG AGC T <u>TGG</u>
	Off-target	AA TTG TAAG TGC TCAG AGC T T <u>*AG</u>

Avoid guides with perfect off-target matches (adjacent to NGG or NAG PAM)

	On-target	AA TTG TAAG TGC TCAG AGC T <u>TGG</u>
	Off-target	AA <u>TGG</u> TTAG TGC TCAG AGC T TGG
Better	On-target	AA TTG TAAG TGC TCAG AGC T <u>TGG</u>
	Off-target	AA TTG TAAG TGC TCAG TGC C TGG

Prioritize guides that have no off-targets with perfectly matching seed regions

On-target	AA TTG TAAG TGC TCAG AGC T <u>TGG</u>
Off-target	G A TGG TAA A TGC TCAG AGC T TGG

Better	On-target	AA TTG TAAG TGC TCAG AGC T <u>TGG</u>
	Off-target	AA TGG TAA A TGC TCAG TGC T TGG

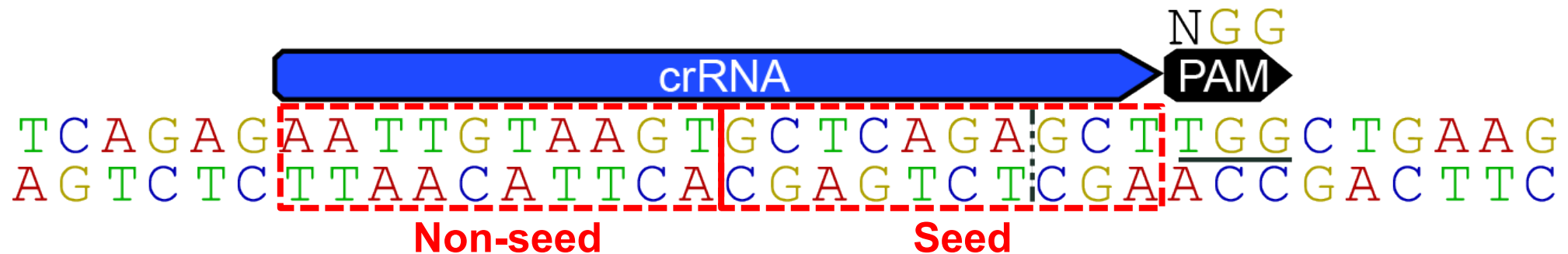
Prioritize guides with off-targets w/ more than 2 mismatches

Better	On-target	AA TTG TAAG TGC TCAG AGC T <u>TGG</u>
	Off-target	AA TTG TAA A TGC TCAG TGC C TGG



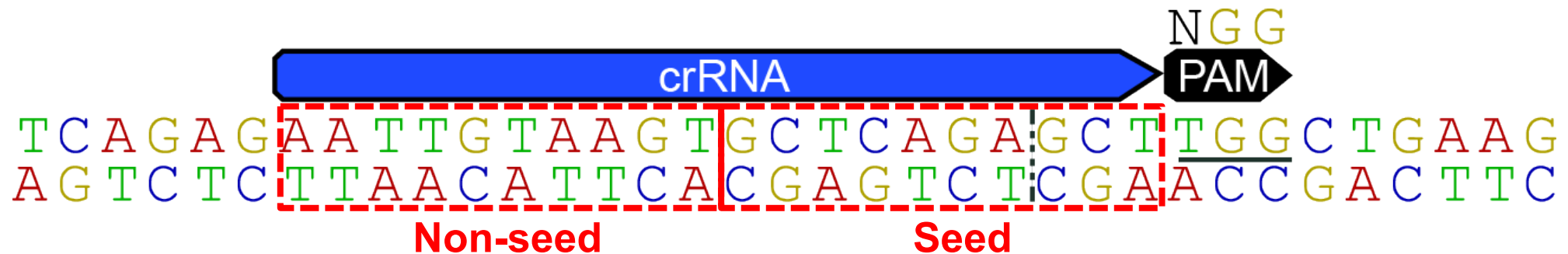
Editing design: Selecting CRISPR guides

- Select guides by balancing:
 - Off-targets
 - Cleavage efficiency



Editing design: Selecting CRISPR guides

- Select guides by balancing:
 - Off-targets
 - Cleavage efficiency
 - Distance from insertion site

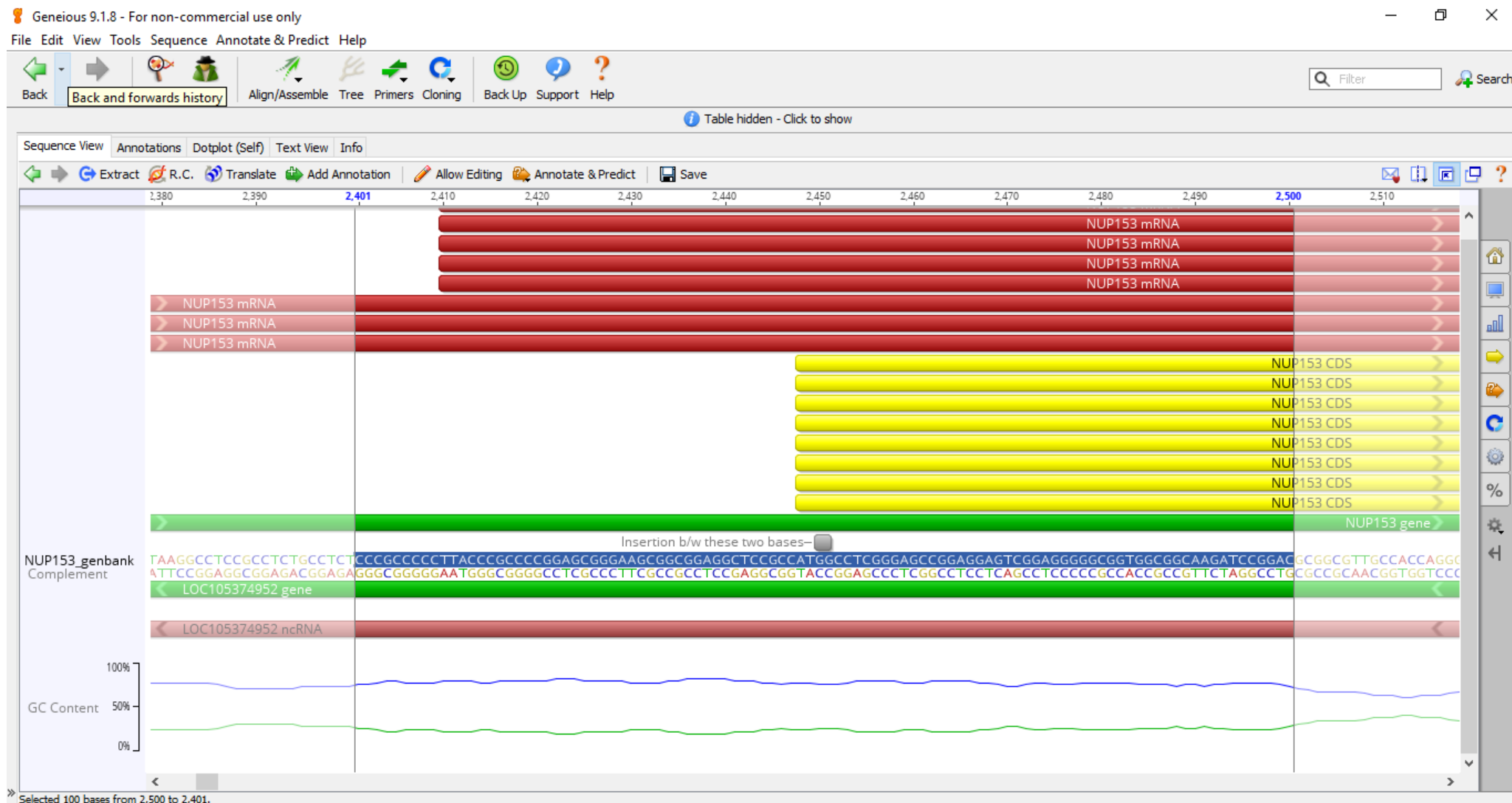


Editing design: Selecting CRISPR guides

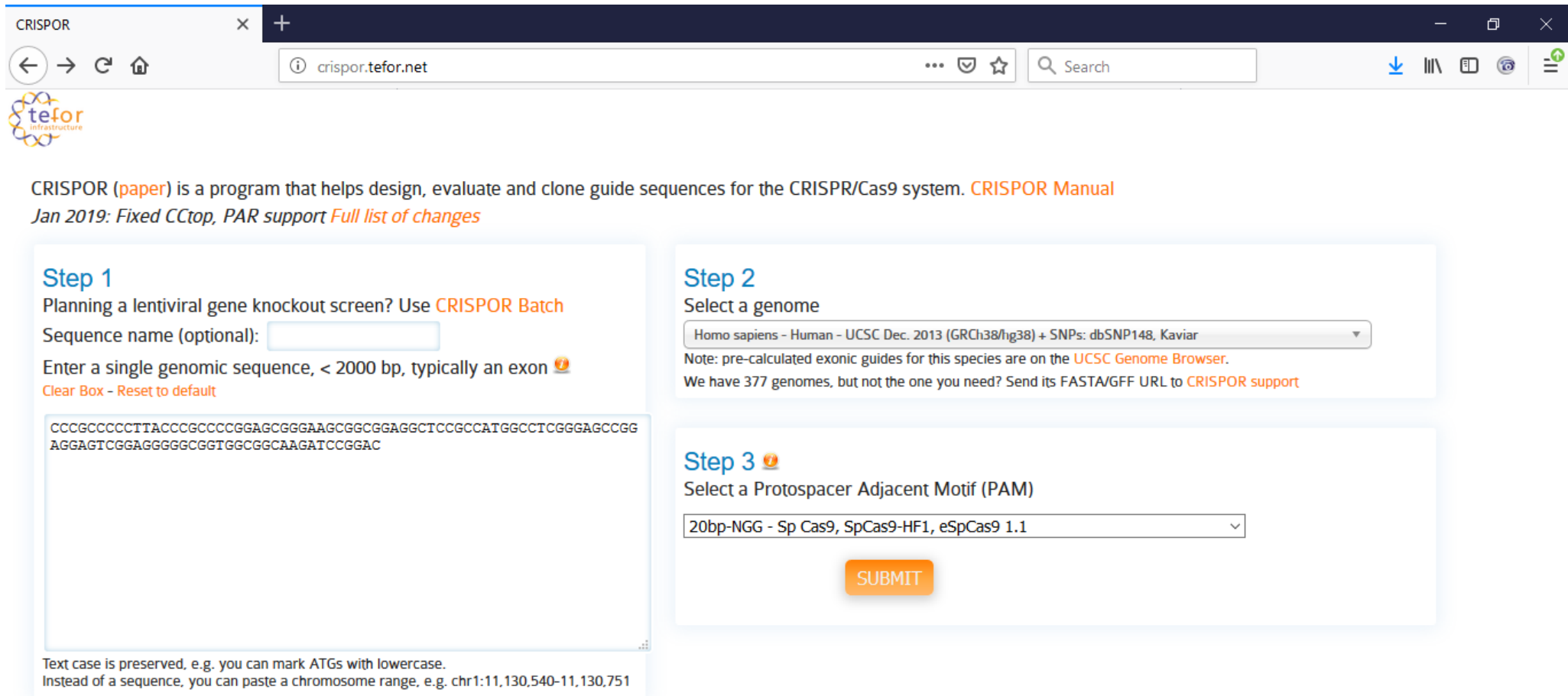
- There are **MANY** tools for CRISPR guide design tools
 - https://en.wikipedia.org/wiki/CRISPR/Cas_Tools
- Some useful tools:
 - CRISPOR
 - UCSC genome browser pre-computed CRISPR tracks for human and mouse



Editing design: Selecting guides w/ CRISPOR



Editing design: Selecting guides w/ CRISPOR



The screenshot shows the CRISPOR web application interface. At the top is a browser window with the address bar showing 'crispor.tefor.net'. Below the browser is the CRISPOR logo. The main content area is divided into three steps:

- Step 1**: Planning a lentiviral gene knockout screen? Use [CRISPOR Batch](#). It includes a text input for 'Sequence name (optional):', a text input for 'Enter a single genomic sequence, < 2000 bp, typically an exon' with a help icon, and a 'Clear Box - Reset to default' link. A text area contains the sequence: `CCCGCCCCCTTACCCGCCCCGGAGCGGGAAGCGGCGGAGGCTCCGCCATGGCCTCGGGAGCCGGAGGAGTCGGAGGGGGCGGTGGCGGCAAGATCCGGAC`.
- Step 2**: Select a genome. A dropdown menu shows 'Homo sapiens - Human - UCSC Dec. 2013 (GRCh38/hg38) + SNPs: dbSNP148, Kaviar'. Below it is a note: 'Note: pre-calculated exonic guides for this species are on the [UCSC Genome Browser](#). We have 377 genomes, but not the one you need? Send its FASTA/GFF URL to [CRISPOR support](#)'.
- Step 3**: Select a Protospacer Adjacent Motif (PAM). A dropdown menu shows '20bp-NGG - Sp Cas9, SpCas9-HF1, eSpCas9 1.1'. Below the dropdown is an orange 'SUBMIT' button.

At the bottom of the interface, there is a footer with the text: 'Text case is preserved, e.g. you can mark ATGs with lowercase. Instead of a sequence, you can paste a chromosome range, e.g. chr1:11,130,540-11,130,751'.

Version 4.7 - [Documentation](#) - [Contact us](#) - [Downloads/local installation](#) - [Paper](#) - [License](#)

<http://crispor.tefor.net/>



Editing design: Selecting guides w/ CRISPOR

CRISPOR: hg38

crispor.tefor.net/crispor.py?batchId=eR4LRfwZYWMgiaeyxvat

tefor infrastructure

Homo sapiens (hg38), chr6:17706335-17706434, reverse genomic strand

Found 34 possible guide sequences in input (100 bp). Click on a PAM NGG match to show its 20 bp guide sequence.
Shown below are the PAM site and the expected cleavage position located -3bp 5' of the PAM site.
Colors **green**, **yellow** and **red** indicate high, medium and low specificity of the PAM's guide sequence in the genome.

Variant database: dbSNP 148 Common SNPs Update Missing a variant database? We can probably add it.

Position	0	10	20	30	40	50	60	70	80	90
Sequence	CCCCGCCCCCTTACCCGCCCCGGAGCGGGAAGCGGCGGAGGCTCCGCCATGGCCTCGGGAGCCGGAGGAGTCGGAGGGGGCGGTGGCGGCAAGATCCGGAC									
Guides	CCC---	CCC---	---CGG	---CGG	CCG---	CCT---	CCG---	---AGG	---TGG	---CGG
	CCG---	CCG---	---GGG	---AGG	CCA---	---CGG	---CGG	---CGG	---	---
	CCC---	CCC---	---CGG	---	---TGG	---	---AGG	---GGG	---	---CGG
	CCC---	CCC---	---	---	---CGG	---	---	---GGG	---	---
	CCC---	CCG---	---	---	---	---	---	---	---	---
	CCT---	---	---	---	---	---	---	---	---	---

Download for: [SerialCloner \(free\)](#) - [ApE \(free\)](#) - [GenomeCompiler](#) - [Benchling](#) - [SnapGene](#) - [Geneious](#) - [Vector NTI](#) - [LaserGene](#) - [Genbank](#) - [FASTA](#) - [Copy sequence to clipboard](#)

Predicted guide sequences for PAMs

Ranked by default from highest to lowest specificity score ([Hsu et al., Nat Biot 2013](#)). Click on a column title to rank by a score.
If you use this website, please cite our [CRISPOR paper in Gen Biol 2016](#). Too much information? Look at the [CRISPOR manual](#).

Download as Excel tables: [Guides](#) / [Off-targets](#) / [Saturating mutagenesis assistant](#)

Position/ Strand	Guide Sequence + PAM + Restriction Enzymes + Variants	Specificity Score	Predicted Efficiency Show all scores	Out-of- Frame score	Off-targets for 0-1-2-3-4 mismatches	Genome Browser links to matches sorted by CFD off-target score <input type="checkbox"/> exons only <input type="checkbox"/> chr6 only
---------------------	---	----------------------	---	------------------------	--	---



Editing design: Selecting guides w/ CRISPOR

CRISPOR

crispor.tefor.net/crispor.py?batchId=eR4LRfWZYWMgiaeyxvat&showAllSco

Download as Excel tables: [Guides](#) / [Off-targets](#) / [Saturating mutagenesis assistant](#)

Position/ Strand	Guide Sequence + PAM + Restriction Enzymes + Variants <input type="checkbox"/> Only G- <input type="checkbox"/> Only GG- <input type="checkbox"/> Only A-	Specificity Score	Predicted Efficiency Main scores Doench '16 Doench '16-Old Chari Xu Doench '14 Wang Moreno-Mateos Azimuth in-vitro CCTop	Out- of- Frame	Off-targets for 0-1-2-3-4 mismatches + next to PAM	Genome Browser links to matches sorted by CFD off-target score <input type="checkbox"/> exons only <input type="checkbox"/> chr6 only
8 / rev	TTCCCGCTCCGGGGCGGGTA AGG Cloning / PCR primers	94	34 29 48 -0.40 25 63 53 64	59	0-0-0-1-51 0-0-0-0-0 52 off-targets	4:intergenic:TSPAN6-SRPX2 4:intergenic:RP11-544P1.3-USP6NL 4:intergenic:APOBEC3A-APOBEC3B show all...
7 / rev	TCCCGCTCCGGGGCGGGTAA GGG Cloning / PCR primers	92	38 35 12 -0.25 20 54 54 81	59	0-0-0-4-50 0-0-0-1-2 54 off-targets	4:intron:CH507-9B2.4/CH507-9B2.3 4:intron:C21orf33 4:intergenic:PHF2-MIR4291 show all...
6 / rev	CCCCTCCGGGGCGGGTAAG GGG ⚠ High GC content Cloning / PCR primers	88	33 28 83 -0.22 26 72 52 69	58	0-0-2-5-72 0-0-1-0-1 79 off-targets	4:intron:MAP3K15 4:exon:ITGAV 4:intergenic:BIRC6-TTC27 show all...
5 / rev	CCGCTCCGGGGCGGGTAAG GGG ⚠ High GC content Cloning / PCR primers	85	38 36 94 0.6 1 49 86 68 72	56	0-0-0-10-95 0-0-0-0-0	3:intron:MOV10 4:intergenic:RP11-796E10.1-AC106900.6 4:exon:KCNQ5 show all...



Editing design: Selecting guides w/ CRISPOR

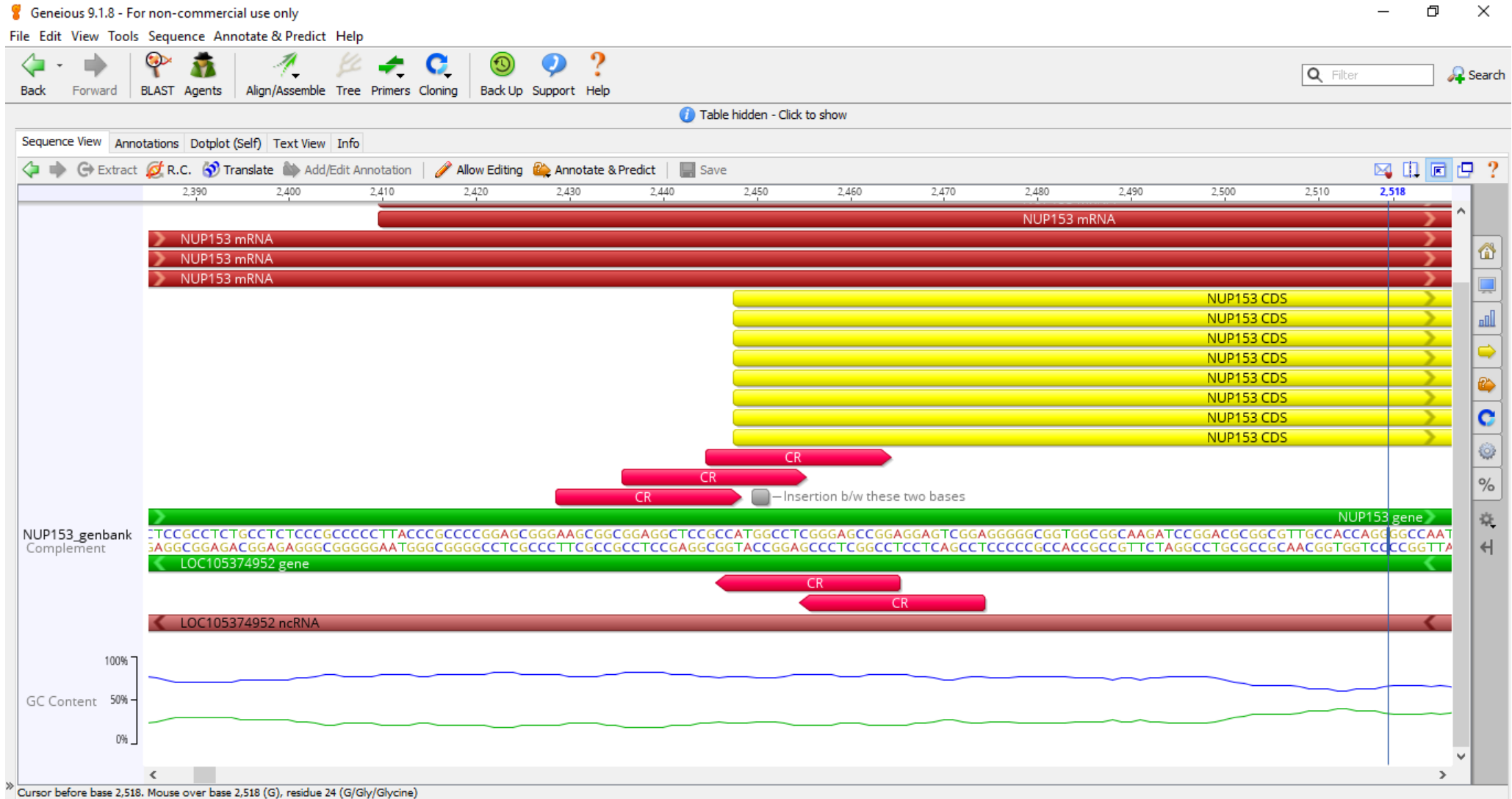
Download as Excel tables: [Guides](#) / [Off-targets](#) / [Saturating mutagenesis assistant](#)

Position/ Strand ⓘ	Guide Sequence + PAM + Restriction Enzymes ⓘ + Variants ⓘ <input type="checkbox"/> Only G- <input type="checkbox"/> Only GG- <input type="checkbox"/> Only A- ⓘ	Specificity Score ⓘ	Predicted Efficiency ⓘ Main scores	Out- of- Frame ⓘ Click score for details	Off-targets for 0-1-2-3-4 mismatches + next to PAM ⓘ	Genome Browser links to matches sorted by CFD off-target score ⓘ <input type="checkbox"/> exons only <input type="checkbox"/> chr6 only
8 / rev	TTCCCGCTCCGGGGCGGGTA AGG Cloning / PCR primers	94	Doench '16 Doench '16-Old Chari Xu Doench '14 Wang Moreno-Mateos Azimuth in-vitro CCTop	59	0-0-0-1-51 0-0-0-0-0 52 off-targets	4:intergenic:TSPAN6-SRPX2 4:intergenic:RP11-544P1.3-USP6NL 4:intergenic:APOBEC3A-APOBEC3B 4:intron:TEAD3 4:intergenic:SLIT2-SLIT2-IT1 4:intergenic:TRMT61A-RNU7-160P 4:intergenic:RP4-630A11.3-LEPR 4:exon:LRR4B 4:intron:TAB2 4:intron:PKN3 4:exon:LINC00957 4:intergenic:RP11-16N2.1-LPHN3 4:intron:FAM129A 4:exon:PIGP 4:exon:CACNA1C-AS1/CACNA1C 4:intergenic:PAWR-PPP1R12A 4:intergenic:FUCA2-PHACTR2 4:intergenic:LINC01326-RN7SKP298 4:intergenic:XKR4-RP11-628E19.2 4:intergenic:ATP6V1D/EIF2S1-EIF2S1 4:intergenic:UBE2I-BAIAP3 4:intron:SPATA17

genome.ucsc.edu/cgi-bin/hqTracks?db=hq38&position=chrX:100639314-100639336



Editing design: Selecting guides w/ CRISPOR



Outline

- Intro to the Allen Institute for Cell Science
- Gene editing design
 - Introduction
 - crRNA design
 - **Donor template design**
- Workflows and methods for generating high-quality clonal hiPS cell lines
- In-lab demonstrations
- Roundtable discussion



Basic Overview for Donor Molecule Design

N-terminal tag



C-terminal tag



What you'll need

- ❑ Gene locus file (showing DNA sequence) with crRNA target sites annotated (we are using **NUP153**)
- ❑ Sequence of FP of choice (we will use **mEGFP**)
- ❑ Amino acid sequence of linker (we are using **YSDLE**)
- ❑ Codon frequency table for your species (we are using **human**)
- ❑ Link to UCSC Genome Browser (with the “conservation” track enabled)



Human codon frequency table

Triplet	Amino acid	Fraction	Frequency/ Thousand	Number	Triplet	Amino acid	Fraction	Frequency/ Thousand	Number
TTT	F	0.45	16.9	336562	TCT	S	0.18	14.6	291040
TTC	F	0.55	20.4	406571	TCC	S	0.22	17.4	346943
TTA	L	0.07	7.2	143715	TCA	S	0.15	11.7	233110
TTG	L	0.13	12.6	249879	TCG	S	0.06	4.5	89429
TAT	Y	0.43	12.0	239268	TGT	C	0.45	9.9	197293
TAC	Y	0.57	15.6	310695	TGC	C	0.55	12.2	243685
TAA	*	0.28	0.7	14322	TGA	*	0.52	1.3	25383
TAG	*	0.20	0.5	10915	TGG	W	1.00	12.8	255512
CTT	L	0.13	12.8	253795	CCT	P	0.28	17.3	343793
CTC	L	0.20	19.4	386182	CCC	P	0.33	20.0	397790
CTA	L	0.07	6.9	138154	CCA	P	0.27	16.7	331944
CTG	L	0.41	40.3	800774	CCG	P	0.11	7.0	139414
CAT	H	0.41	10.4	207826	CGT	R	0.08	4.7	93458
CAC	H	0.59	14.9	297048	CGC	R	0.19	10.9	217130
CAA	Q	0.25	11.8	234785	CGA	R	0.11	6.3	126113
CAG	Q	0.75	34.6	688316	CGG	R	0.21	11.9	235938
ATT	I	0.36	15.7	313225	ACT	T	0.24	12.8	255582
ATC	I	0.48	21.4	426570	ACC	T	0.36	19.2	382050
ATA	I	0.16	7.1	140652	ACA	T	0.28	14.8	294223
ATG	M	1.00	22.3	443795	ACG	T	0.12	6.2	123533
AAT	N	0.46	16.7	331714	AGT	S	0.15	11.9	237404
AAC	N	0.54	19.5	387148	AGC	S	0.24	19.4	385113
AAA	K	0.42	24.0	476554	AGA	R	0.20	11.5	228151
AAG	K	0.58	32.9	654280	AGG	R	0.20	11.4	227281
GTT	V	0.18	10.9	216818	GCT	A	0.26	18.6	370873
GTC	V	0.24	14.6	290674	GCC	A	0.40	28.5	567930
GTA	V	0.11	7.0	138154	GCA	A	0.23	16.0	317338
GTG	V	0.47	28.9	575438	GCG	A	0.11	7.6	150708
GAT	D	0.46	22.3	443369	GGT	G	0.16	10.8	215544
GAC	D	0.54	26.0	517579	GGC	G	0.34	22.8	453917
GAA	E	0.42	29.0	577846	GGA	G	0.25	16.3	325243
GAG	E	0.58	40.8	810842	GGG	G	0.25	16.4	326879



Human codon frequency table

Triplet	Amino acid	Fraction	Frequency/ Thousand	Number	Triplet	Amino acid	Fraction	Frequency/ Thousand	Number
TTT	F	0.45	16.9	336562	TCT	S	0.18	14.6	291040
TTC	F	0.55	20.4	406571	TCC	S	0.22	17.4	346943
TTA	L	0.07	7.2	143715	TCA	S	0.15	11.7	233110
TTG	L	0.13	12.6	249879	TCG	S	0.06	4.5	89429
TAT	Y	0.43	12.0	239268	TGT	C	0.45	9.9	197293
TAC	Y	0.57	15.6	310695	TGC	C	0.55	12.2	243685
TAA	*	0.28	0.7	14322	TGA	*	0.52	1.3	25383
TAG	*	0.20	0.5	10915	TGG	W	1.00	12.8	255512
CTT	L	0.13	12.8	253795	CCT	P	0.28	17.3	343793
CTC	L	0.20	19.4	386182	CCC	P	0.33	20.0	397790
CTA	L	0.07	6.9	138154	CCA	P	0.27	16.7	331944
CTG	L	0.41	40.3	800774	CCG	P	0.11	7.0	139414
CAT	H	0.41	10.4	207826	CGT	R	0.08	4.7	93458
CAC	H	0.59	14.9	297048	CGC	R	0.19	10.9	217130
CAA	Q	0.25	11.8	234785	CGA	R	0.11	6.3	126113
CAG	Q	0.75	34.6	688316	CGG	R	0.21	11.9	235938
ATT	I	0.36	15.7	313225	ACT	T	0.24	12.8	255582
ATC	I	0.48	21.4	426570	ACC	T	0.36	19.2	382050
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GTC	V	0.24	14.6	290874	GCC	A	0.40	28.5	567930
GTA	V	0.11	7.0	139156	GCA	A	0.23	16.0	317338
GTG	V	0.47	28.9	575438	GCG	A	0.11	7.6	150708
GAT	D	0.46	22.3	443369	GGT	G	0.16	10.8	215544
GAC	D	0.54	26.0	517579	GGC	G	0.34	22.8	453917
GAA	E	0.42	29.0	577846	GGA	G	0.25	16.3	325243
GAG	E	0.58	40.8	810842	GGG	G	0.25	16.4	326879



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Human BLAT Search

BLAT Search Genome

Genome: ☐ Search ALL Assembly: Query type: Sort output: Output type:

Human Dec. 2013 (GRCh38/hg38) BLAT's guess query.score hyperlink

GGGCCGGGCGGCGGGGAGGCTCTCTAAGGCTCCGCCTCTGCCTCTCCCGCCCCCTTACCCGCCCCGGAGCGGGAAG
CGGCGGAGGCTCCGCCATG

submit I'm feeling lucky clear

Paste in a query sequence to find its location in the the genome. Multiple sequences may be searched if separated by lines starting with '>' followed by the sequence name.

File Upload: Rather than pasting a sequence, you can choose to upload a text file containing the sequence.
Upload sequence:

Choose File No file chosen submit file

Only DNA sequences of 25,000 or fewer bases and protein or translated sequence of 10000 or fewer letters will be processed. Up to 25 sequences can be submitted at the same time. The total limit for multiple sequence submissions is 50,000 bases or 25,000 letters.
A valid example is `GTCTCGGAACCAGGACCTCGGCGTGGCCTAGCG` (human SOD1).

The **Search ALL** checkbox above the Genome drop-down list allows you to search the genomes of the default assemblies for all of our organisms. It also searches any attached hubs' blat servers. This shows you which organisms have the highest homology with your query sequence. The results are ordered so that the organism whose best alignment has the most hits is at the top, and shows the best region found. It makes quick approximate alignments based only on the raw hits, which are a perfectly matching short sub-sequence of a fixed size: 11 for DNA and 4 for protein. The entire alignment, including mismatches and gaps, must score 20 or higher in order to appear in the BLAT output. Having too few hits will often yield no BLAT results. Click the Assembly column link on the results page to see the full BLAT output for that organism.

For locating PCR primers, use [In-Silico PCR](#) for best results instead of BLAT.

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Human (hg38) BLAT Results

BLAT Search Results

Go back to [chr6:17706385-17706445](#) on the Genome Browser.

Custom track name:

Custom track description:

[Build a custom track with these results](#)

ACTIONS	QUERY	SCORE	START	END	QSIZE	IDENTITY	CHROM	STRAND	START	END	SPAN
browser details	YourSeq	97	1	97	97	100.0%	chr6	-	17706385	17706481	97
browser details	YourSeq	26	59	94	97	96.6%	chr19	+	590724	590761	38
browser details	YourSeq	23	31	54	97	100.0%	chr9	-	137760135	137760162	28
browser details	YourSeq	21	30	50	97	100.0%	chr16	+	71588821	71588841	21

[Missing a match?](#)

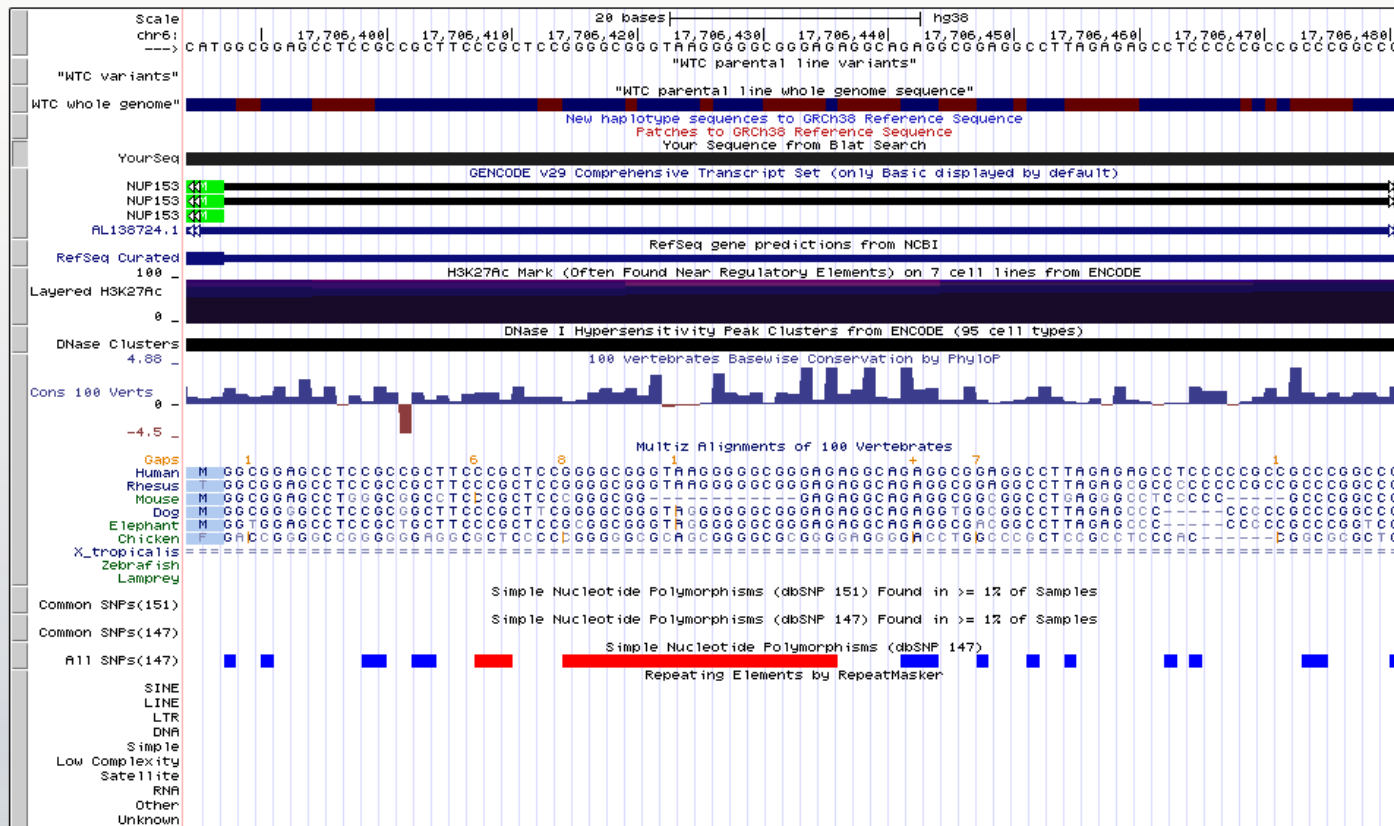


UCSC Genome Browser on Human Dec. 2013 (GRCh38/hg38) Assembly

move <<< << < > >> >>> zoom in 1.5x 3x 10x base zoom out 1.5x 3x 10x 100x

chr6:17,706,385-17,706,481 97 bp.

chr6 (p22.3) 6p22.3 6q126q13q14.1 q1516.1 6q21 22.31 266q27



move start

< 2.0 >

Click on a feature for details. Click or drag in the base position track to zoom in. Click side bars for track options. Drag side bars or labels up or down to reorder tracks. Drag tracks left or right to new position. Press "?" for keyboard shortcuts.

move end

< 2.0 >

track search

default tracks

default order

ide all add cu

Custom tracks

track hubs

configure

multi-regional

reverse

resize re

resh

collapse all

Use drop-down controls below and press refresh to alter tracks displayed. Tracks with lots of items will automatically be displayed in more compact modes.

expand all

WTC genome

disconnect

refresh



hide

hide

hide

-

mRNA and EST

refresh

P12

Human mRNAs

P12

Spliced ESTs

P12

Human ESTs

P12

Other ESTs

P12

Other mRNAs

SIB

Alt-Splicing

hide

hide

hide

hide

hide

hide

-

Expression

refresh

19

P12

GTEX Gene

GTEX Transcript

Affy_GNF1H

Affy_U133

Affy_U95

GNF Atlas 2

hide

hide

hide

hide

hide

hide

GWIPS-viz Riboseq

miRNA Tissue Atlas

hide

hide

-

Regulation

refresh

19

P12

ENCODE Regulation...

Updated

GeneHancer

P12

CpG Islands...

ORegAnno

RefSeq Func Elems

show

hide

hide

hide

hide

-

Comparative Genomics

refresh

Conservation

Cons 7 Verts

Cons 20 Mammals

Cons 30 Primates

Primate Chain/Net

Placental Chain/Net

full

hide

hide

hide

hide

hide

Vertebrate Chain/Net

Chicken Chain/Net

hide

hide

-

Variation

refresh

Common SNPs(151)

Common SNPs(150)

Common SNPs(147)

Common SNPs(146)

Common SNPs(144)

Common SNPs(142)

dense

hide

dense

hide

hide

hide

Common SNPs(141)

All SNPs(151)

All SNPs(150)

All SNPs(147)

All SNPs(146)

All SNPs(144)

hide

hide

hide

dense

hide

hide

All SNPs(142)

All SNPs(141)

Flagged SNPs(151)

Flagged SNPs(150)

Flagged SNPs(147)

Flagged SNPs(146)

hide

hide

hide

hide

hide

hide

Flagged SNPs(144)

Flagged SNPs(142)

Flagged SNPs(141)

Mult. SNPs(151)

Mult. SNPs(150)

Mult. SNPs(147)

hide

hide

hide

hide

hide

hide

Mult. SNPs(146)

Mult. SNPs(144)

Mult. SNPs(142)

Mult. SNPs(141)

DGV Struct Var

hide

hide

hide

hide

hide

-

Repeats

refresh

P12

RepeatMasker

Interrupted Rpts

RepeatMasker Viz.

Microsatellite

Segmental Dups

Self Chain



UCSC Genome Browser on Human Dec. 2013 (GRCh38/hg38) Assembly

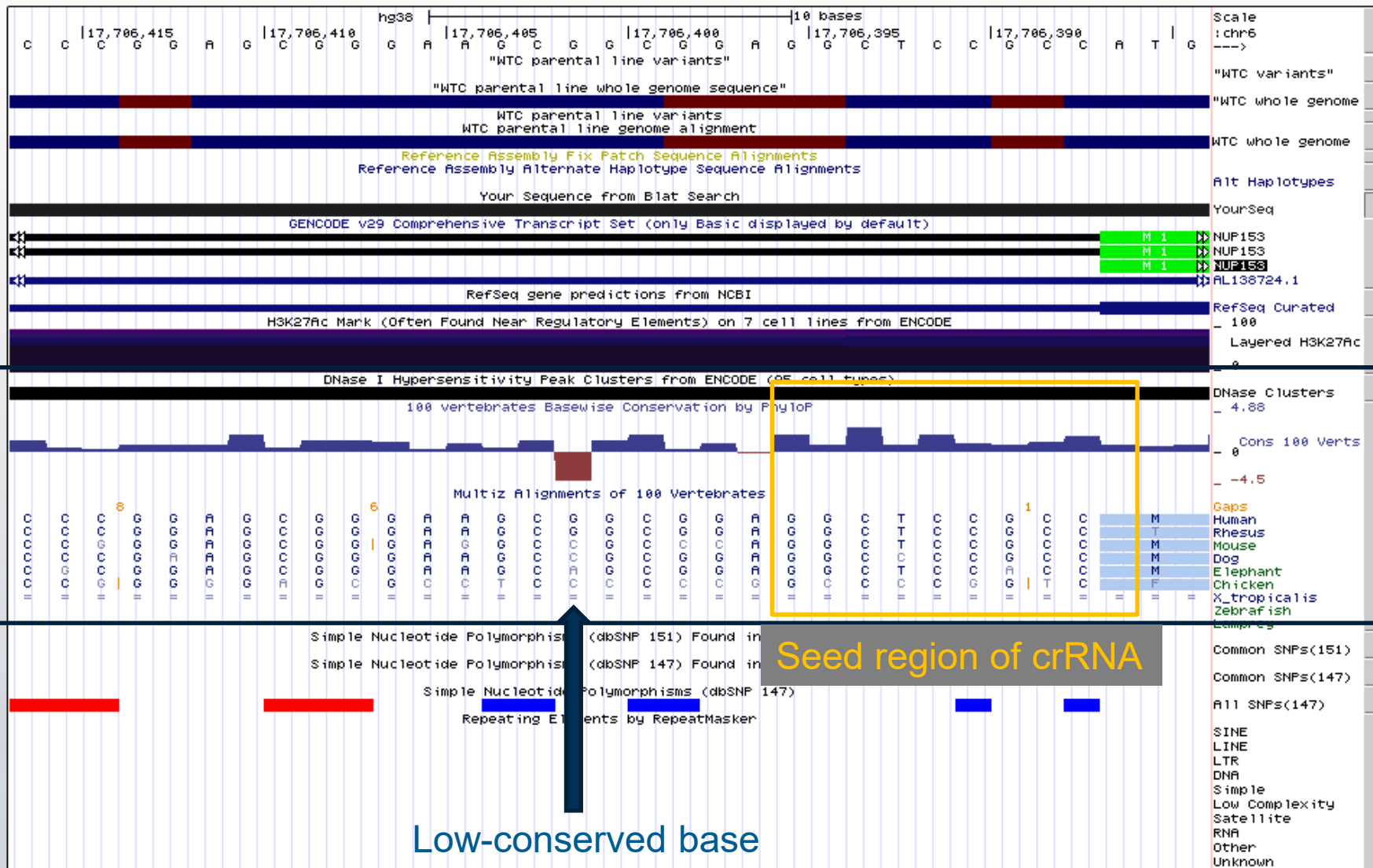
move <<< << < > >> >>> zoom in 1.5x 3x 10x base zoom out 1.5x 3x 10x 100x

chr6:17,706,385-17,706,417 33 bp. enter position, gene symbol, HGVS or search terms



go

6q27.26 22.31 6q21 16.1q15 14.1q13.6q12 6p22.3 chr6 (p22.3)

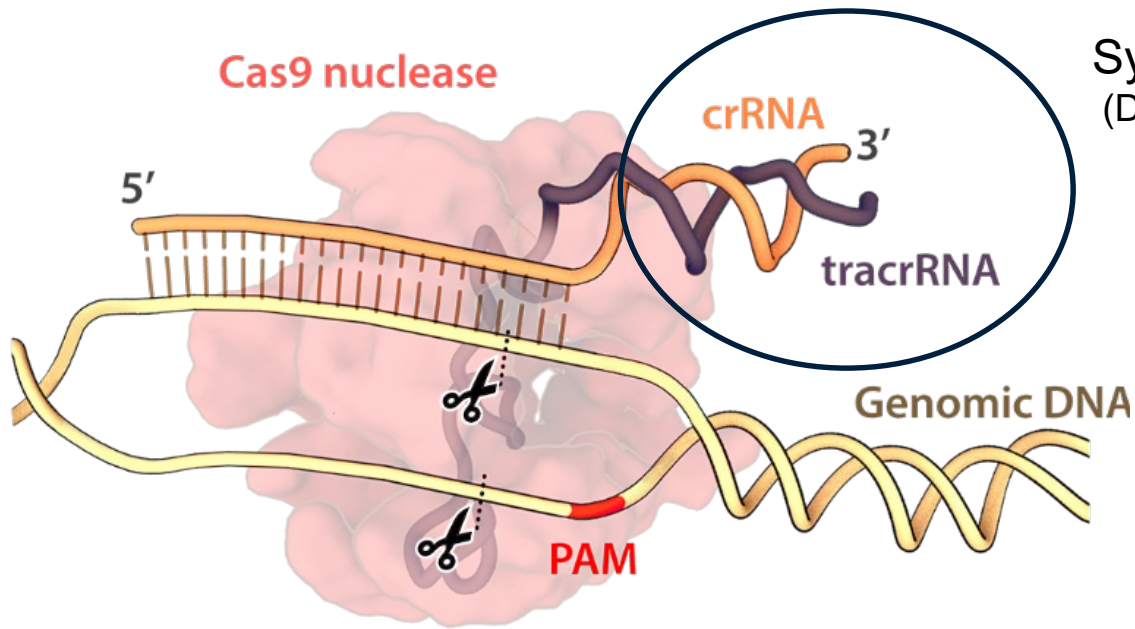


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GAG	E	0.58	40.8	810842	GGG	G	0.25	16.4	326879



Ordering Reagents: crRNAs and Donor Templates

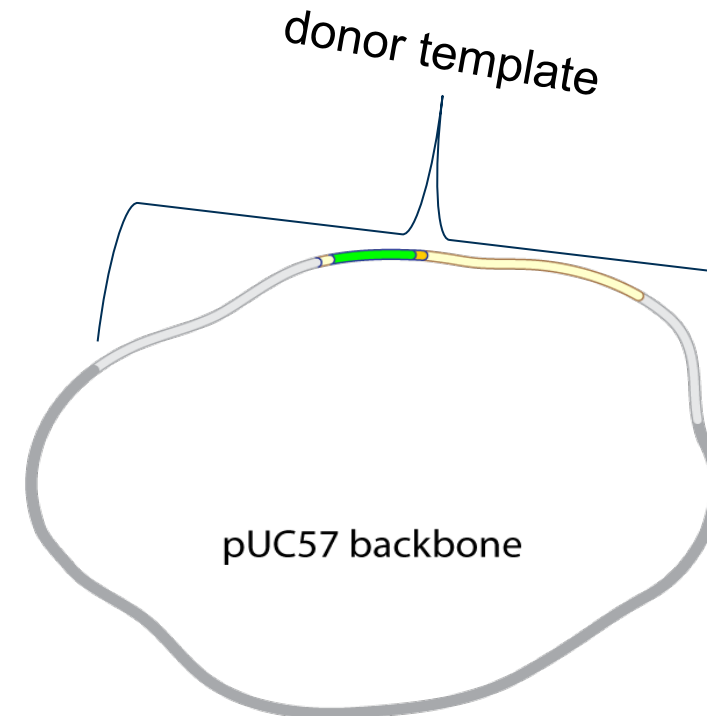


Synthetic unmodified 2-part crRNA and tracrRNA

(Dharmacon, Horizon Discovery)

Pros: short-lived in cells, easy to order, pure, fast turnaround time

Cons: have to purchase tracrRNA, too



Plasmid, synthesized commercially

(Genewiz)

Pros: easy to order, sequence-verified, customizable backbone

Cons: costly, takes 6-8 weeks, can randomly integrate in genome, can be toxic to cells



Next!

